

MAY 20 2003

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(Translation)

Mailed: April 15, 2003

NOTIFICATION OF REASONS FOR REJECTION

RECEIVED

Patent Application No.: 2000-247729

MAY 22 2003

Examiner's Notice Date: April 8, 2003

TECH CENTER 1600/2900

Examiner: Keiko Nagai

This application is rejected on the grounds stated below. Any opinion about the rejection must be filed within 60 DAYS of the mailing date hereof.

REASON

The invention is unpatentable under Section 29 (2) of the Patent Law, as being such that the invention could easily have been made by a person with ordinary skill in the art to which the invention pertains, on the basis of the invention described in the following publication(s) distributed in Japan or a foreign country prior to this application.

REMARKS

(1) Claim 1 : Reference 1

Reference 1 discloses a nucleotide sequence of an estrogen receptor gene of medaka fish. Here, it is easily achievable for a person having ordinary skill in the art to prepare a probe based on the nucleotide sequence of the gene discussed in Reference 1, thus obtaining polynucleotide including upstream and downstream regions of the estrogen receptor gene of medaka fish and determining its nucleotide sequence.

(2) Claim 2 : Reference 1

Reference 1 discloses a nucleotide sequence of an estrogen receptor gene of medaka fish. Here, it is easily achievable for a person having ordinary

discussed in Reference 1, thus obtaining an estrogen receptor gene of medaka fish and determining its nucleotide sequence.

(3) Claim 3 : Reference 1

Reference 1 discloses a nucleotide sequence of an estrogen receptor gene of medaka fish, and an amino acid sequence of the estrogen receptor. Here, it is easily achievable for a person having ordinary skill in the art to prepare a probe based on the nucleotide sequence of the gene discussed in Reference 1, thus obtaining an estrogen receptor gene of medaka fish and determining its nucleotide sequence and an amino acid sequence encoded by it.

(4) Claim 4 : Reference 1

As described in remarks (1) and (2) above, it is easily achievable for a person having ordinary skill in the art to obtain polynucleotide recited in Claims 1 and 2. Assembly of a recombinant vector is merely a well-known means.

(5) Claim 5 : Reference 1

Introduction of a gene to a host in order to examine an in-vivo function of a product of an isolated gene is well-known means.

The claims not mentioned in this Official Action are not rejected. If a new reason for rejection is noticed, a further Official Action will be issued.

Reference Cited:

1. Jpn. Pat. Appln. KOKAI Publication 2000-201688

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Prior Art Search Report

Searched Field: IPC 7th ed. C12N 15/00

SwissProt/PIR/GeneSeq

Genbank/EMBL/DDBJ/GeneSeq

BIOSIS

MEDLINE

WPIDS

Prior-Art Document(s):

Winn R., Marine Environmental Research, vol. 46 (1-5), p.130 (1998)

pp. 192-199 (1994)

Gray M.A. et al., Environmental Toxicology and Chemistry, vol. 18(11), pp. 2587-2594 (1999)

The result of this prior art search does not constitute the reasons for rejection.

Mailing Date: April 15, 2005

整理番号 A 0 0 0 0 0 3 8 8 5

発送番号 1 2 1 2 6 4  
発送日 平成 1 5 年 4 月 1 5 日

1 / 3

## 拒絶理由通知書

特許出願の番号	特願 2 0 0 0 - 2 4 7 7 2 9
起案日	平成 1 5 年 4 月 8 日
特許庁審査官	長井 啓子 9 1 2 3 4 N 0 0
特許出願人代理人	鈴江 武彦 (外 5 名) 様
適用条文	第 2 9 条第 2 項

15.6.14

この出願は、次の理由によって拒絶をすべきものである。これについて意見があれば、この通知書の発送の日から 6 0 日以内に意見書を提出して下さい。

### 理 由

この出願の下記の請求項に係る発明は、その出願前日本国内又は外国において頒布された下記 of 刊行物に記載された発明に基いて、その出願前にその発明の属する技術の分野における通常の知識を有する者が容易に発明をすることができたものであるから、特許法第 2 9 条第 2 項の規定により特許を受けることができない。

記 (引用文献等については引用文献等一覧参照)

#### (1) 請求項 1 : 引用文献 1

引用文献 1 には、メダカのエストロゲンレセプター遺伝子の塩基配列が開示されている。引用文献 1 記載の遺伝子の塩基配列を基にしてプローブを作成して、メダカのエストロゲンレセプター遺伝子上流及び下流の領域を含むポリヌクレオチドを得てその塩基配列を決定することは、当業者が容易になし得る程度のことにすぎない。

#### (2) 請求項 2 : 引用文献 1

引用文献 1 には、メダカのエストロゲンレセプター遺伝子の塩基配列が開示されている。引用文献 1 記載の遺伝子の塩基配列を基にしてプローブを作成して、メダカのエストロゲンレセプター遺伝子を得てその塩基配列を決定することは、当業者が容易になし得る程度のことにすぎない。

#### (3) 請求項 3 : 引用文献 1

引用文献 1 には、メダカのエストロゲンレセプター遺伝子の塩基酸配列及び当該エストロゲンレセプターのアミノ酸配列が開示されている。引用文献 1 記載の

広島大学長

遺伝子の塩基配列を基にしてプローブを作成して、メダカのエストロゲンレセプター遺伝子を得てその塩基配列及びそれがコードするアミノ酸配列を決定すること、当業者が容易になし得る程度のことにはすぎない。

(4) 請求項4：引用文献1

請求項1及び請求項2記載のポリヌクレオチドを得ることが、引用文献1の記載に基づいて当業者が容易になし得たことは、上記(1)及び(2)で説明したとおりである。組み換えベクターを構築することは常套手段にすぎない。

(5) 請求項5：引用文献1

単離した遺伝子の産物の生体内機能を探る等の目的で、宿主に遺伝子導入することは、常套手段である。

この拒絶理由通知書中で指摘した請求項以外の請求項に係る発明については、現時点では、拒絶の理由を発見しない。拒絶の理由が新たに発見された場合には拒絶の理由が通知される。

引用文献等一覧

1. 特開2000-201688号公報

この拒絶理由通知書に不明な点がある場合、または、この案件について面接を希望する場合は、

特許審査第三部生命工学 長井 啓子

Tel. 03-3581-1101 (特許庁代表)

Fax. 03-3501-0491

までご連絡下さい。

先行技術文献調査結果の記録

・調査した分野 IPC第7版 C12N 15/00  
SwissProt/PIR/GeneSeq  
Genbank/EMBL/DDBJ/GeneSeq

MEDELIAL

WPIDS

ID AAA92174 standard; DNA; 1728 BP.  
XX  
AC AAA92174; XP-002181484  
XX

DT 05-JAN-2001 (first entry)  
XX

DE Oryzias lapites oestrogen receptor encoding DNA SEQ ID NO:2.  
XX

KW Oryzias lapites; oestrogen receptor; ds.  
XX

OS Oryzias lapites.  
XX

PN JP2000201688-A.  
XX

PD 25-JUL-2000.  
XX

PF 06-APR-1999; 99JP-0098787.  
XX

PR 10-NOV-1998; 98JP-0319465.  
XX

PA (SUMO ) SUMITOMO CHEM CO LTD.  
XX

DR WPI; 2000-567950/53.  
DR

DR P-PSDB; AAB20897.  
XX

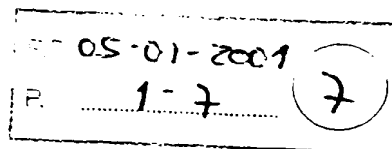
PT An estrogen receptor gene and its application -  
XX

PS Claim 3; Page 11-13; 23pp; Japanese.  
XX

CC The present sequence encodes an oestrogen receptor derived from  
CC Oryzias lapites. Also described are: (1) a vector comprising the  
CC oestrogen receptor gene; (2) a transformant prepared by introducing  
CC the oestrogen receptor gene or vector from (1) into a host cell;  
CC (3) a method for the preparation of an oestrogen receptor comprising  
CC culturing the transformant from (2) to produce the oestrogen receptor;  
CC and (4) a method for the evaluation of oestrogen receptor-activating  
CC ability of a chemical substance in which the chemical substance is  
CC reacted with a transformant prepared by introducing a reporter gene  
CC connected downstream of a transcription controlling region containing  
CC an oestrogen response sequence and the above oestrogen receptor gene to  
CC an oestrogen-nonendogenous host cell. The transformant can be used for  
CC the evaluation of oestrogen receptor-activating ability of a chemical  
CC substance.  
XX

SQ Sequence 1728 BP; 378 A; 514 C; 497 G; 339 T; 0 other;

atgtaccctg	aagagagccg	gggttctgga	ggggtggctg	ctgtggacct	tttggaaagg	60
acgtacgact	atgccgcccc	caaccctgcc	acgactcccc	tttacagcca	gtccagcacc	120
ggctactact	ctgctcccc	ggaacaaac	ggacccccct	cagaaggcag	tctgcagtcc	180
ctgggcagtg	ggccgacgag	ccctctggtg	tttgtgccct	ccagccccag	actcagtccc	240
tttatgcatc	caccagcca	ccactatctg	gaaaccactt	ccacgcccgt	ttacagatcc	300
agccaccagg	gagcctccag	ggaggaccag	tgcggctccc	gggaggacac	gtgcagcctg	360
ggggagttag	gcgccggagc	cggggctggg	gggtttgaga	tggccaaaga	cacgcgtttc	420
tgcgccgtgt	gcagcgacta	cgctctggtg	taccactatg	gggtgtggtc	ttgtgagggc	480
tgcaaggcct	tcttcaagag	gagcatccag	ggtcacaatg	actatatgtg	cccagcgacc	540
aatcagtgca	ctattgacag	aaatcgaagg	aagggctgtc	aggcttgtcg	tcttaggaag	600
tgttacgaag	tgggaatgat	gaaaggcggt	gtgcgcgaag	accgcattcg	cattttacgg	660
cgtgacaaac	ggcggacagg	cgttggtgat	ggagacaagg	ttgtaaagg	tcaggagcat	720
aaaacggtgc	attatgatgg	aaggaaacgc	agcagcacag	gaggaggagg	aggaggagga	780
ggaggaagac	tgtctgtgac	cagcatacct	cctgagcagg	tgctgtctct	ccttcagggc	840
gccgagcccc	cgatactctg	ctcgcgtcag	aagttgagcc	gaccgtacac	cgaggtcacc	900
atgatgacct	tgctcaccag	catggcagac	aaggagctgg	tccacatgat	cgcctgggcc	960
aagaagctcc	cagggtttct	gcagctgtcc	ctgcacgata	aggtgctgct	gctggagagc	1020
tcgtggctgg	aggtgctcat	gatcggcctc	atttggaggt	ccatccactg	tcccgggaag	1080
ctcatctttg	cacaagacct	catcctggac	aggaatgagg	gagactgcgt	ggaaggcatg	1140



acggagatct	togacatgct	gctggccact	gcttcccgt	tccgtgtgct	caaactcaaa	1200
cctgaggaat	tcgtctgct	caaagctatt	attttactca	actccggtgc	tttttctttc	1260
tgacccggca	ccatggagcc	acttcacaac	<del>agcgcggcgg</del>	ttcagagcat	gctggacacc	1320
atcacagacg	cactcattca	ttacatcagt	cagtcgggtt	acttgccca	ggagcaggcg	1380
agacggcagg	cccagccgt	cctgctgctc	tcccacatca	ggcacatgag	caacaaaggc	1440
atggagcacc	tctacagcat	gaagtgcagg	aacaaagtcc	ctctttatga	cctcctactg	1500
<del>gagatgctcg</del>	<del>atgcccaccg</del>	<del>cctgcaccac</del>	<del>cccgtcagag</del>	<del>ccccccagtc</del>	<del>cttgtcccaa</del>	1560
gtcgacagag	accctccctc	caccagcagc	ggcgggggtg	gaatcgctcc	cggttctata	1620
tcagcatctc	gaggcagaat	<del>cgagagtcgg</del>	<del>agcagaggcc</del>	cctttgctcc	<del>cagtgtcctt</del>	1680
cagtatggag	ggtcgcgtcc	tgactgcacc	ccggcccttc	aagactga		1728

//

>>GSN:AAA92174 Oryzias lapites oestrogen recept (1728 nt)  
 initn: 8586 initl: 8586 opt: 8586 Z-score: 8271.0 bits: 1544.2 E(): 0  
 99.653% identity (99.653% ungapped) in 1728 nt overlap (211-1938:1-1728)

	190	200	210	220	230	240
EP0111	CGCCTCTCGCCCCGTGACCCCTCGGTGACATGTACCCCTGAAGAGAGCCGGGGTTCTGGA					
						.....
GSN:AA			ATGTACCCCTGAAGAGAGCCGGGGTTCTGGA			
			10	20	30	

	250	260	270	280	290	300
EP0111	GGGGTGGCTGCTGTGGACTTTTGGGAAGGGACGTACGACTATGCCGCCCCCAACCCCTGCC					
						.....
GSN:AA	GGGGTGGCTGCTGTGGACCTTTTGGGAAGGGACGTACGACTATGCCGCCCCCAACCCCTGCC					
	40	50	60	70	80	90

	310	320	330	340	350	360
EP0111	ACGACTCCCCCTTACAGCCAGTCCAGCACCGGCTACTACTCTGCTCCCTGGAAACAAAC					
						.....
GSN:AA	ACGACTCCCCCTTACAGCCAGTCCAGCACCGGCTACTACTCTGCTCCCTGGAAACAAAC					
	100	110	120	130	140	150

	370	380	390	400	410	420
EP0111	GGACCCCTCAGAAGGCAGTCTGCAGTCCCTGGGCAGTGGGCCGACGAGCCCTCTGGTG					
						.....
GSN:AA	GGACCCCTCAGAAGGCAGTCTGCAGTCCCTGGGCAGTGGGCCGACGAGCCCTCTGGTG					
	160	170	180	190	200	210

	430	440	450	460	470	480
EP0111	TTTGTGCCCTCCAGCCCCAGACTCAGTCCCTTTATGCATCCACCCAGCCCACTATCTG					
						.....
GSN:AA	TTTGTGCCCTCCAGCCCCAGACTCAGTCCCTTTATGCATCCACCCAGCCCACTATCTG					
	220	230	240	250	260	270

	490	500	510	520	530	540
EP0111	GAAACCACTTCCACGCCCGTTTACAGATCCAGCCACCAGGGAGCCTCCAGGGAGGACCAG					
						.....
GSN:AA	GAAACCACTTCCACGCCCGTTTACAGATCCAGCCACCAGGGAGCCTCCAGGGAGGACCAG					
	280	290	300	310	320	330

	550	560	570	580	590	600
EP0111	TGCGGCTCCCGGGAGGACACGTGCAGCCTGGGGGAGTTAGGCGCCGGAGCCGGGGCTGGG					
						.....
GSN:AA	TGCGGCTCCCGGGAGGACACGTGCAGCCTGGGGGAGTTAGGCGCCGGAGCCGGGGCTGGG					
	340	350	360	370	380	390

	610	620	630	640	650	660
EP0111	GGGTTTGAGATGGCCAAAGACACGCGTTTCTGCGCCGTGTGCAGCGACTACGCCTCTGGG					
						.....
GSN:AA	GGGTTTGAGATGGCCAAAGACACGCGTTTCTGCGCCGTGTGCAGCGACTACGCCTCTGGG					
	400	410	420	430	440	450

	670	680	690	700	710	720
EP0111	TACCACTATGGGGTGTGGTCTTGTGAGGGCTGCAAGGCCTTCTTCAAGAGGAGCATCCAG					
						.....
GSN:AA	TACCACTATGGGGTGTGGTCTTGTGAGGGCTGCAAGGCCTTCTTCAAGAGGAGCATCCAG					
	460	470	480	490	500	510

	730	740	750	760	770	780
EP0111	GGTCACAATGACTATATGTGCCCAGCGACCAATCAGTGCACCTATTGACAGAAATCGGAGG					
						.....
GSN:AA	GGTCACAATGACTATATGTGCCCAGCGACCAATCAGTGCACCTATTGACAGAAATCGAAGG					
	520	530	540	550	560	570





	1450	1460	1470	1480	1490	1500
EP0111	ATTTTACTCAACTCCGGTGCTTTTTCCTTTCTGCAACCGGCAACCATGGAGCCACTTCACAAC					
GSN:AA	ATTTTACTCAACTCCGGTGCTTTTTCCTTTCTGCAACCGGCAACCATGGAGCCACTTCACAAC					
	1240	1250	1260	1270	1280	1290
	1510	1520	1530	1540	1550	1560
EP0111	AGCGCGGCGGTTTCAGAGCATGCTGGACACCATCACAGACGGCACTCATTTCATTACATCAGT					
GSN:AA	AGCGCGGCGGTTTCAGAGCATGCTGGACACCATCACAGACGGCACTCATTTCATTACATCAGT					
	1300	1310	1320	1330	1340	1350
	1570	1580	1590	1600	1610	1620
EP0111	CAGTCGGGTTACTTTGGCCCAGGAGCAGGCGAGACGGCAGGCCAGCTGCTCCTGCTGCTC					
GSN:AA	CAGTCGGGTTACTTTGGCCCAGGAGCAGGCGAGACGGCAGGCCAGGCCGCTCCTGCTGCTC					
	1360	1370	1380	1390	1400	1410
	1630	1640	1650	1660	1670	1680
EP0111	TCCCACATCAGGCACATGAGCAACAAAGGCATGGAGCACCTCTACAGCATGAAGTCCAAAG					
GSN:AA	TCCCACATCAGGCACATGAGCAACAAAGGCATGGAGCACCTCTACAGCATGAAGTCCAAAG					
	1420	1430	1440	1450	1460	1470
	1690	1700	1710	1720	1730	1740
EP0111	AACAAAGTCCCTCTTTATGACCTCCTACTGGAGATGCTCGATGGGCAACCGCCTGCACCAAC					
GSN:AA	AACAAAGTCCCTCTTTATGACCTCCTACTGGAGATGCTCGATGGGCAACCGCCTGCACCAAC					
	1480	1490	1500	1510	1520	1530
	1750	1760	1770	1780	1790	1800
EP0111	CCCGTCAGAGCACCCCAAGTCCTTGTCCCAAGTCGACAGAGACCTCCCTCCACCAGCAGC					
GSN:AA	CCCGTCAGAGCCCCCAAGTCCTTGTCCCAAGTCGACAGAGACCTCCCTCCACCAGCAGC					
	1540	1550	1560	1570	1580	1590
	1810	1820	1830	1840	1850	1860
EP0111	GGCGGGGGTGGGAATCGCTCCCGGTTCTATATCAGCATCTCGAGGCAGAATCGAGAGTCCG					
GSN:AA	GGCGGGGGTGGGAATCGCTCCCGGTTCTATATCAGCATCTCGAGGCAGAATCGAGAGTCCG					
	1600	1610	1620	1630	1640	1650
	1870	1880	1890	1900	1910	1920
EP0111	AGCAGAGGCCCTTTGCTCCCAAGTGTCTTCAGTATGGAGGGTCGCGTCCTGACTGCACC					
GSN:AA	AGCAGAGGCCCTTTGCTCCCAAGTGTCTTCAGTATGGAGGGTCGCGTCCTGACTGCACC					
	1660	1670	1680	1690	1700	1710
	1930	1940	1950	1960	1970	1980
EP0111	CCGGCCCTTCAAGACTGAGCACACAGTCCAAGGCCCTTTTTTGTGGCTCAAGGGTTCAG					
GSN:AA	CCGGCCCTTCAAGACTGA					
	1720					

17  
ID AAB20897 standard; Protein; 575 AA.

XX

AC AAB20897;

XX

DT 05-JAN-2001 (first entry)

XX

DE Oryzias lapites oestrogen receptor protein SEQ ID NO:1.

XX

KW Oryzias lapites; oestrogen receptor.

XX

OS Oryzias lapites.

XX

PN JP2000201688-A.

XX

PD 25-JUL-2000.

XX

PF 06-APR-1999; 99JP-0098787.

XX

PR 10-NOV-1998; 98JP-0319465.

XX

PA (SUMO ) SUMITOMO CHEM CO LTD.

XX

DR WPI; 2000-567950/53.

DR N-PSDB; AAA92174.

XX

PT An estrogen receptor gene and its application

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PS Claim 1; Page 9-10; 23pp; Japanese.

XX

CC The present sequence represents an oestrogen receptor derived from  
CC Oryzias lapites. Also described are: (1) a vector comprising the  
CC oestrogen receptor gene; (2) a transformant prepared by introducing  
CC the oestrogen receptor gene or vector from (1) into a host cell;  
CC (3) a method for the preparation of an oestrogen receptor comprising  
CC culturing the transformant from (2) to produce the oestrogen receptor;  
CC and (4) a method for the evaluation of oestrogen receptor-activating  
CC ability of a chemical substance in which the chemical substance is  
CC reacted with a transformant prepared by introducing a reporter gene  
CC connected downstream of a transcription controlling region containing  
CC an oestrogen response sequence and the above oestrogen receptor gene to  
CC an oestrogen-nonendogenous host cell. The transformant can be used for  
CC the evaluation of oestrogen receptor-activating ability of a chemical  
CC substance.

XX

SQ Sequence- 575 AA;

SQ 37 A; 38 R; 10 N; 27 D; 0 B; 18 C; 24 Q; 29 E; 0 Z; 58 G; 19 H;

SQ 22 I; 61 L; 24 K; 19 M; 14 F; 37 P; 58 S; 29 T; 4 W; 20 Y; 27 V;

SQ 0 Others;

mypeesrgsg gvaavdlleg tydyaapnpa ttplyqsst gyysapletn gppsegsllqs  
lgsqptsplv fvpssprlsp fmhppsghyl ettstpvrys shqgasredq cgsredtcs1  
gelgagagag gfemakdtrf cavcsdyasg yhygvwsceg ckaffkrsiq ghndymcpat  
nqctidrnrr kgcqacrlrk cyevgmkkqg vrkdririlr rdkrrtgvd qdkvvkqgeh  
ktvhydgrkr sstggggggg ggrlsvtsip peqvlllllg aeppilcsrq klsrpytevt  
mmtlltsmad kelvhmiawa kklpgflqls lhdqvlles swlevlmigl iwrshcpqgk  
lifaqdlild rnegdcvegm teifdmllat asrfrvlklk peefvcl kai illnsgafsf  
ctgtmeplhn saavqsmltd itdalihiyis qsgylaqeqa rrqaqpllll shirhmsnkq  
mehlysmkck nkvpdydlld emldahrlbh pvrpqslsq vdrdpstss ggggiapgsi  
sasrgriesp srgpfapsvl qyggsrpdct palqd

//

```
>>GSP:AAB20897 Oryzias latipes oestrogen recept (575 aa)
initn: 3905 initl: 3905 opt: 3905 Z-score: 2879.0 bits: 544.9 E(): 2.1e-152
Smith-Waterman score: 3905; 99.478% identity (99.478% ungapped) in 575 aa overlap
(210-1934:1-575)
```

390            420            450            480            510            540  
 EP0111 LGSGPTSPLVFPSSPRLSPFMHPPSHHYLETTSTPVYRSSHQGASREDQCGSREDTCSL  
 ::  
 GSP:AA LGSGPTSPLVFPSSPRLSPFMHPPSHHYLETTSTPVYRSSHQGASREDQCGSREDTCSL  
           70            80            90            100            110            120

570            600            630            660            690            720  
EP0111 GELGAGAGAGGFEMAKDTRFCVCSGYHYGVWSCEGCKAFFKRSIQGHNDYMCPAT  
         ::  
GSP:AA GELGAGAGAGGFEMAKDTRFCVCSGYHYGVWSCEGCKAFFKRSIQGHNDYMCPAT  
         130            140            150            160            170            180

[illegible]

930            960            990            1020            1050            1080  
EP0111 KTVHYDGRKRSSSTGGGGGGGGGRLSVTSIPPEQVLLLLLQGAEPPIILCSRQKLSRPYTEVT  
         :         :         :         :         :         :         :         :         :  
GSP:AA KTVHYDGRKRSSSTGGGGGGGGGRLSVTSIPPEQVLLLLLQGAEPPIILCSRQKLSRPYTEVT  
         250            260            270            280            290            300

```

      1110      1140      1170      1200      1230      1260
EP0111 MMTLLTSMADKELVHMIAWAKKLPGLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGK
      :
      :
      :
GSP:AA MMTLLTSMADKELVHMIAWAKKLPGLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGK
      310      320      330      340      350      360

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      1290      1320      1350      1380      1410      1440
EP0111 LIFAQDLILDRNEGDCVEGMTEIFDMLLATASRFRVLKCLKPEEFVCLKAIILLNSGAFS
      :
      :
      :
GSP:AA LIFAQDLILDRNEGDCVEGMTEIFDMLLATASRFRVLKCLKPEEFVCLKAIILLNSGAFS
      370      380      390      400      410      420

```

```

      1470      1500      1530      1560      1590      1620
EP0111 CTGTMEPLHNSAAVQSMLDTITDALIHYISQSGYLAQEQARRQAQLLLLLSHIRHMSNKG
      :         :         :         :         :         :
GSP:AA CTGTMEPLHNSAAVQSMLDTITDALIHYISQSGYLAQEQARRQAQLLLLLSHIRHMSNKG
           430           440           450           460           470           480

```

```

      1650      1680      1710      1740      1770      1800
EP0111 MEHLYSMKCKNKVPLYDLLLEMLDAHRLHHPVRAPQSLSQVDRDPPSTSSGGGGIAPGSI
      :
      :
      :
GSP:AA MEHLYSMKCKNKVPLYDLLLEMLDAHRLHHPVRAPQSLSQVDRDPPSTSSGGGGIAPGSI
      490      500      510      520      530      540

```

```

      1830      1860      1890      1920
EP0111 SASRGRIESPSRGPFPASVLQYGGSRPDCTPALQD
      : : : : : : : : : : : : : : : : : : : :
GSP:AA SASRGRIESPSRGPFPASVLQYGGSRPDCTPALQD
           550           560           570

```

ID ' AAA92175 standard; DNA; 1863 BP.

XX

AC AAA92175;

XP-002181483

XX

DT 05-JAN-2001 (first entry)---

XX

DE Oryzias lapites oestrogen receptor encoding DNA-SEQ-ID NO:4. -

XX

KW Oryzias lapites; oestrogen receptor, ds.

XX

OS Oryzias lapites.

XX

PN JP2000201688-A.

XX

PD 25-JUL-2000.

XX

PF 06-APR-1999; 99JP-0098787.

XX

PR 10-NOV-1998; 98JP-0319465.

XX

PA (SUMO ) SUMITOMO CHEM CO LTD.

XX

DR WPI; 2000-567950/53.

DR

P-PSDB; AAB20898.

XX

PT An estrogen receptor gene and its application -

XX

PS Claim 4; Page 15-17; 23pp; Japanese.---

XX

CC The present sequence encodes an oestrogen receptor derived from  
CC Oryzias lapites. Also described are: (1) a vector comprising the  
CC oestrogen receptor gene; (2) a transformant prepared by introducing  
CC the oestrogen receptor gene or vector from (1) into a host cell;  
CC (3) a method for the preparation of an oestrogen receptor comprising  
CC culturing the transformant from (2) to produce the oestrogen receptor;  
CC and (4) a method for the evaluation of oestrogen receptor-activating  
CC ability of a chemical substance in which the chemical substance is  
CC reacted with a transformant prepared by introducing a reporter gene  
CC connected downstream of a transcription controlling region containing  
CC an oestrogen response sequence and the above oestrogen receptor gene to  
CC an oestrogen-nonendogenous host cell. The transformant can be used for  
CC the evaluation of oestrogen receptor-activating ability of a chemical  
CC substance.

XX

SQ Sequence 1863 BP; 406 A; 565 C; 531 G; 361 T; 0 other;

atgagtaaga gacagagctc ggtgcagatc aggcagctgt tcggaccagc actcagatcc	60
aggatcagcc cagcctcctc agagctggag accctctccc cacctcgctt ctgccccgt	120
gacccccctcg gtagaatgta cctgaagag agccggggtt ctggaggggt ggctgctgtg	180
gaccttttgg aagggacgta cgactatgcc gccccaacc ctgccacgac tcccccttac	240
agccagtgca gacccggcta ctactctgt cccctggaaa caaacggacc cccctcagaa	300
ggcagtcctgc agtccctggg cagtgggccc acgagccctc tgggtgttgc gccctccagc	360
cccagactca gtccctttat gcatccaccc agccaccact atctggaaac caattccacg	420
cccgtttaca gatccagcca ccaggaggcc tccaggaggg accagtgcgg ctcccgggag	480
gacacgtgca gcctggggga gttaggcgcc ggagccgggg ctgggggggt tgagatggcc	540
aaagacacgc gtttctgcgc cgtgtgcagc gactacgcct ctgggtacca ctatgggggtg	600
tggctctgtg agggctgcaa ggccttcttc aagaggagca tccagggtca caatgactat	660
atgtgcccag cgaccaatca gtgcactatt gacagaaatc gaaggaaggg ctgtcaggct	720
tgtcgtctta ggaagtgtta cgaagtggga atgatgaaag gcggtgtgcg caaggaccgc	780
attcgcatth tacggcgtga caaacggcgg acaggcggtg gtgatggaga caaggttgta	840
aagggtcagg agcataaaac ggtgcattat gatggaagga aacgcagcag cacaggagga	900
ggaggaggag gaggaggagg aagactgtct gtgaccagca tacctcctga gcaggtgctg	960
ctcctccttc agggcgccga gccccgata ctctgctcgc gtcagaagtt gagccgaccg	1020
tacaccgagg tcacatgat gaccctgctc accagcatgg cagacaagga gctggtccac	1080
atgatcgctt gggccaagaa gctcccaggt tttctgcagc tgtccctgca cgatcagggt	1140

ctgctgctgg	agagctcgtg	gctggaggtg	ctcatgatcg	gcctcatttg	gaggtccatc	1200
cactgtcccg	ggaagctcat	ctttgcacaa	gacctcatcc	tggacaggaa	tgagggagac	1260
tgcgtggaag	gcatgacgga	gatcttcgac	atgctgctgg	ccactgcttc	ccgcttcogt	1320
gtgctcaaac	tcaaacctga	ggaattcgtc	tgctcaaag	ctattatatt	actcaactcc	1380
ggtgcttttt	ctttctgcac	cggcaccatg	gagccacttc	acaacagcgc	ggcggttcag	1440
agcatgctgg	acaccatcac	agacgcactc	attcattaca	tcagtcagtc	gggttacttg	1500
gcccaggagc	aggcgagacg	gcaggcccag	ccgctcctgc	tgctctccca	catcaggcac	1560
atgagcaaca	aaggcatgga	gcacctctac	agcatgaagt	gcaagaacaa	agtccctctt	1620
tatgacctcc	tactggagat	gctcgatgcc	caccgcctgc	accaccccg	cagagccccc	1680
cagtccttgt	cccaagtcga	cagagaccct	ccctccacca	gcagcggcgg	gggtggaatc	1740
gctcccgggt	ctatatcagc	atctcgaggc	agaatcgaga	gtccgagcag	aggccctttt	1800
gctcccagtg	tccttcagta	tggaggggtc	cgtcctgact	gcaccccggc	ccttcaagac	1860
tga						1863

//

>>GSN:AAA92175 Oryzias lapites oestrogen recept (1863 nt)  
initn: 9261 initl: 9261 opt: 9261 Z-score: 8922.0 bits: 1664.7 E(): 0  
99.678% identity (99.678% ungapped) in 1863 nt overlap (76-1938:1-1863)

```
      50      60      70      80      90     100
EP0111 CGTGTTCGCGCAGCACATCTGAGCATGATTCATGAGTAAGAGACAGAGCTCGGTGCAGATC
      :
GSN:AA      ATGAGTAAGAGACAGAGCTCGGTGCAGATC
                  10      20      30
```

```
     110     120     130     140     150     160
EP0111 AGGCAGCTGTTTCGGACCAGCACTCAGATCCAGGATCAGCCCAGCCTCCTCAGAGCTGGAG
      :
GSN:AA AGGCAGCTGTTTCGGACCAGCACTCAGATCCAGGATCAGCCCAGCCTCCTCAGAGCTGGAG
          40      50      60      70      80      90
```

```
     170     180     190     200     210     220
EP0111 ACCCTCTCCCCACCTCGCCTCTCGCCCCGTGACCCCCCTCGGTGACATGTACCTGAAGAG
      :
GSN:AA ACCCTCTCCCCACCTCGCCTCTCGCCCCGTGACCCCCCTCGGTGACATGTACCTGAAGAG
          100     110     120     130     140     150
```

```
     230     240     250     260     270     280
EP0111 AGCCGGGGTTTCTGGAGGGGTGGCTGCTGTGGACTTTTGGAAAGGGACGTACGACTATGCC
      :
GSN:AA AGCCGGGGTTTCTGGAGGGGTGGCTGCTGTGGACTTTTGGAAAGGGACGTACGACTATGCC
          160     170     180     190     200     210
```

```
     290     300     310     320     330     340
EP0111 GCCCCCAACCCTGCCACGACTCCCTTTACAGCCAGTCCAGCACCGGCTACTACTCTGCT
      :
GSN:AA GCCCCCAACCCTGCCACGACTCCCTTTACAGCCAGTCCAGCACCGGCTACTACTCTGCT
          220     230     240     250     260     270
```

```
     350     360     370     380     390     400
EP0111 CCCCTGGAAACAAACGGACCCCCCTCAGAAGGCAGTCTGCAGTCCCTGGGCAGTGGGCGG
      :
GSN:AA CCCCTGGAAACAAACGGACCCCCCTCAGAAGGCAGTCTGCAGTCCCTGGGCAGTGGGCGG
          280     290     300     310     320     330
```

```
     410     420     430     440     450     460
EP0111 ACGAGCCCTCTGGTGTGTTGTGCCCTCCAGCCCCAGACTCAGTCCCTTTATGCATCCACCC
      :
GSN:AA ACGAGCCCTCTGGTGTGTTGTGCCCTCCAGCCCCAGACTCAGTCCCTTTATGCATCCACCC
          340     350     360     370     380     390
```

```
     470     480     490     500     510     520
EP0111 AGCCACCACTATCTGGAAACCACTTCCACGCCCCGTTTACAGATCCAGCCACCAGGGAGCC
      :
GSN:AA AGCCACCACTATCTGGAAACCACTTCCACGCCCCGTTTACAGATCCAGCCACCAGGGAGCC
          400     410     420     430     440     450
```

```
     530     540     550     560     570     580
EP0111 TCCAGGGAGGACCACTGCGGCTCCCGGGAGGACACGTGCAGCCTGGGGGAGTTAGGCGCC
      :
GSN:AA TCCAGGGAGGACCACTGCGGCTCCCGGGAGGACACGTGCAGCCTGGGGGAGTTAGGCGCC
          460     470     480     490     500     510
```

```
     590     600     610     620     630     640
EP0111 GGAGCCGGGGCTGGGGGGTTTGAGATGGCCAAAGACACGCGTTTCTGCGCCGTGTGCAGC
      :
GSN:AA GGAGCCGGGGCTGGGGGGTTTGAGATGGCCAAAGACACGCGTTTCTGCGCCGTGTGCAGC
          520     530     540     550     560     570
```

```
     650     660     670     680     690     700
```

BNSDOCID: YP 2181483A 1



	1310	1320	1330	1340	1350	1360
EP0111	GACCTCATCTGGACAGGAATGAGGGAGACTGCGTGGAAGGCATGACGGAGATCTTCGAC					
	::					
GSN:AA	GACCTCATCTGGACAGGAATGAGGGAGACTGCGTGGAAGGCATGACGGAGATCTTCGAC					
	1240	1250	1260	1270	1280	1290

	1370	1380	1390	1400	1410	1420
EP0111	ATGCTGCTGGCCACTGCTTCCCGCTTCCGTGTGCTCAAACCTCAAACCTGAGGAATTCGTC					
	::					
GSN:AA	ATGCTGCTGGCCACTGCTTCCCGCTTCCGTGTGCTCAAACCTCAAACCTGAGGAATTCGTC					
	1300	1310	1320	1330	1340	1350

	1430	1440	1450	1460	1470	1480
EP0111	TGCCTCAAAGCTATTATTTTACTCAACTCCGGTGCTTTTTTCTTCTGCACCGGCACCATG					
	::					
GSN:AA	TGCCTCAAAGCTATTATTTTACTCAACTCCGGTGCTTTTTTCTTCTGCACCGGCACCATG					
	1360	1370	1380	1390	1400	1410

	1490	1500	1510	1520	1530	1540
EP0111	GAGCCACTTCACAACAGCGCGGGTTCAGAGCATGCTGGACACCATCACAGACGCACTC					
	::					
GSN:AA	GAGCCACTTCACAACAGCGCGGGTTCAGAGCATGCTGGACACCATCACAGACGCACTC					
	1420	1430	1440	1450	1460	1470

	1550	1560	1570	1580	1590	1600
EP0111	ATTCATTACATCAGTCAGTCGGGTTACTTGGCCAGGAGCAGGCGAGACGGCAGGCCAG					
	::					
GSN:AA	ATTCATTACATCAGTCAGTCGGGTTACTTGGCCAGGAGCAGGCGAGACGGCAGGCCAG					
	1480	1490	1500	1510	1520	1530

	1610	1620	1630	1640	1650	1660
EP0111	CTGCTCCTGCTGCTCTCCACATCAGGCACATGAGCAACAAAGGCATGGAGCACCTCTAC					
	: ::					
GSN:AA	CCGCTCCTGCTGCTCTCCACATCAGGCACATGAGCAACAAAGGCATGGAGCACCTCTAC					
	1540	1550	1560	1570	1580	1590

	1670	1680	1690	1700	1710	1720
EP0111	AGCATGAAGTGCAAGAACAAAGTCCCTCTTTATGACCTCCTACTGGAGATGCTCGATGCC					
	::					
GSN:AA	AGCATGAAGTGCAAGAACAAAGTCCCTCTTTATGACCTCCTACTGGAGATGCTCGATGCC					
	1600	1610	1620	1630	1640	1650

	1730	1740	1750	1760	1770	1780
EP0111	CACCGCCTGCACCACCCCGTCAGAGCACCCAGTCCTTGTCCCAAGTCGACAGAGACCCT					
	::					
GSN:AA	CACCGCCTGCACCACCCCGTCAGAGCACCCAGTCCTTGTCCCAAGTCGACAGAGACCCT					
	1660	1670	1680	1690	1700	1710

	1790	1800	1810	1820	1830	1840
EP0111	CCCTCCACCAGCAGCGCGGGGTGGAATCGCTCCCGGTTCTATATCAGCATCTCGAGGC					
	::					
GSN:AA	CCCTCCACCAGCAGCGCGGGGTGGAATCGCTCCCGGTTCTATATCAGCATCTCGAGGC					
	1720	1730	1740	1750	1760	1770

	1850	1860	1870	1880	1890	1900
EP0111	AGAATCGAGAGTCCGAGCAGAGGCCCTTTGCTCCCAAGTGTCTTCAAGTATGGAGGGTCG					
	::					
GSN:AA	AGAATCGAGAGTCCGAGCAGAGGCCCTTTGCTCCCAAGTGTCTTCAAGTATGGAGGGTCG					
	1780	1790	1800	1810	1820	1830

	1910	1920	1930	1940	1950	1960
EP0111	CGTCTGACTGCACCCCGGCCCTTCAAGACTGAGCACACAGTCCAAGGCCCTTTTTTTTGT					
	::					
GSN:AA	CGTCTGACTGCACCCCGGCCCTTCAAGACTGA					
	1840	1850	1860			

1970 1980 1990 2000 2010 2020  
EP0111 GGCTCAAGGGTTCAGGTTGGGACAAGGTGATGCTTGATTTAATTTTAAGAATTATTATA

1 Yell.  
translation of Sep 21 1  
VS  
Seq TAB of SP: 20168

ID AAB20898 standard; Protein; 620 AA.  
XX  
AC AAB20898;  
XX  
DT 05-JAN-2001 (first entry)  
XX  
DE Oryzias lapites oestrogen receptor protein SEQ ID NO:3.  
XX  
KW Oryzias lapites; oestrogen receptor.  
XX  
OS Oryzias lapites.  
XX  
PN JP2000201688-A.  
XX  
PD 25-JUL-2000.  
XX  
PF 06-APR-1999; 99JP-0098787.  
XX  
PR 10-NOV-1998; 98JP-0319465.  
XX  
PA (SUMO ) SUMITOMO CHEM CO LTD.  
XX  
DR WPI; 2000-567950/53.  
DR N-PSDB; AAA92175.  
XX  
PT An estrogen receptor gene and its application  
XX  
PS Claim 2; Page 13-15; 23pp; Japanese.  
XX

CC The present sequence represents an oestrogen receptor derived from  
CC Oryzias lapites. Also described are: (1) a vector comprising the  
CC oestrogen receptor gene; (2) a transformant prepared by introducing  
CC the oestrogen receptor gene or vector from (1) into a host cell;  
CC (3) a method for the preparation of an oestrogen receptor comprising  
CC culturing the transformant from (2) to produce the oestrogen receptor;  
CC and (4) a method for the evaluation of oestrogen receptor-activating  
CC ability of a chemical substance in which the chemical substance is  
CC reacted with a transformant prepared by introducing a reporter gene  
CC connected downstream of a transcription controlling region containing  
CC an oestrogen response sequence and the above oestrogen receptor gene to  
CC an oestrogen-nonendogenous host cell. The transformant can be used for  
CC the evaluation of oestrogen receptor-activating ability of a chemical  
CC substance.

XX  
SQ Sequence 620 AA;  
SQ 39 A; 44 R; 10 N; 29 D; 0 B; 18 C; 27 Q; 31 E; 0 Z; 60 G; 19 H;  
SQ 24 I; 67 L; 25 K; 20 M; 15 F; 43 P; 67 S; 30 T; 4 W; 20 Y; 28 V;  
SQ 0 Others;  
mskrqssvqi rqlfgpalrs rispassele tisprrlspr dplgdmypee srgsggvaav  
dllegtydya apnpattply ~~sqsstgyysa pletnppse qslqslgsgp tsplvfvss~~  
prlspfmhpp shhylettst pyvrsshqga sredqgsre dtcslgelga gagaggfema  
kdtrfcavcs dyasgyhygv ~~wscecgckaff krsigghndy mcpatnqcti dmrrkqcca~~  
crlrkcyevg mmkggvrkdr ~~irilrrdkrr tgvqgdgkvv kqgehktvhy dgrkrsstgg~~  
gggggggrls vtsippeqvl ~~lllqgaoppi lcarqqlarp ytevtmtll tsmadkelvb~~  
miawakkllpg flqlslhdqv lllesswlev lmigliwrsi hcpgklifaq dlildrnegd  
cvegmteifd mllatasrfr ~~vklkpeefv cikarillns gsfstctgtm eplhmsaavq~~  
smltditdal ihyisqgyl aqeqarrqaq pllllshirh msnkgmehly smkcknkvpl  
ydlillemda hrlhhpvrp ~~qslsqvdrdp pstssggggi apgsisasrg riespsrgpf~~  
apsvlqyggs rpdctpalqd

//

>>GSP:AAB20898 Oryzias latipes oestrogen recept (620 aa)  
 initn: 4198 initl: 4198 opt: 4198 Z-score: 3093.7 bits: 584.7 E(): 2.3e-164  
 Smith-Waterman score: 4198; 99.516% identity (99.516% ungapped) in 620 aa overlap  
 (75-1934:1-620)

```

      90      120      150      180      210      240
EP0111 MSKRQSSVQIRQLFGPALRSRISPASSELETLSPPRLSPRDPLGDMYPFESRGSGGVAAV
      .....
GSP:AA MSKRQSSVQIRQLFGPALRSRISPASSELETLSPPRLSPRDPLGDMYPFESRGSGGVAAV
      10      20      30      40      50      60

      270      300      330      360      390      420
EP0111 DFLEGTIDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGLQSLGSGPTSPLVFPSS
      .....
GSP:AA DLLEGTIDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGLQSLGSGPTSPLVFPSS
      70      80      90      100      110      120

      450      480      510      540      570      600
EP0111 PRLSPFMHPPSHHYLETTSTPVYRSSHQGASREDQCGSREDTCSLGELGAGAGAGGFEMA
      .....
GSP:AA PRLSPFMHPPSHHYLETTSTPVYRSSHQGASREDQCGSREDTCSLGELGAGAGAGGFEMA
      130      140      150      160      170      180

      630      660      690      720      750      780
EP0111 KDTRFCAVCSYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPATNQCTIDNRNRKSCQA
      .....
GSP:AA KDTRFCAVCSYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPATNQCTIDNRNRKSCQA
      190      200      210      220      230      240

      810      840      870      900      930      960
EP0111 CRLRKCYEVGMMKGGVRKDRIRILRRDKRRTGVGDGDKVVKQEHKTVHYDGRKRSSTGG
      .....
GSP:AA CRLRKCYEVGMMKGGVRKDRIRILRRDKRRTGVGDGDKVVKQEHKTVHYDGRKRSSTGG
      250      260      270      280      290      300

      990      1020      1050      1080      1110      1140
EP0111 GGGGGGGRLSVTSIPPEQVLLLLQGAEPFILCSRQKLSRPYTEVTMTLLTSMADKELVH
      .....
GSP:AA GGGGGGGRLSVTSIPPEQVLLLLQGAEPFILCSRQKLSRPYTEVTMTLLTSMADKELVH
      310      320      330      340      350      360

      1170      1200      1230      1260      1290      1320
EP0111 MIAWAKKLPGLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGKLIFAQDLILDRNEGD
      .....
GSP:AA MIAWAKKLPGLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGKLIFAQDLILDRNEGD
      370      380      390      400      410      420

      1350      1380      1410      1440      1470      1500
EP0111 CVEGMTEIFDMLLATASRFRVLKLPKEEFVCLKAIILLNSGAFSFCGTGTMEPLHNSAAVQ
      .....
GSP:AA CVEGMTEIFDMLLATASRFRVLKLPKEEFVCLKAIILLNSGAFSPCTGTGTMEPLHNSAAVQ
      430      440      450      460      470      480

      1530      1560      1590      1620      1650      1680
EP0111 SMLDTITDALIHYSISQGYLAQEQAARQAQLLLLLSHIRHMSNKGMEHLYSMKCKNKVPL
      .....
GSP:AA SMLDTITDALIHYSISQGYLAQEQAARQAQLLLLLSHIRHMSNKGMEHLYSMKCKNKVPL
      490      500      510      520      530      540

      1710      1740      1770      1800      1830      1860
EP0111 YDLLLEMLDAHRLHHPVRAPQSLSQVDRDPPSTSSGGGGIAPGSISASRGRIESPSRGPF
      .....
GSP:AA YDLLLEMLDAHRLHHPVRAPQSLSQVDRDPPSTSSGGGGIAPGSISASRGRIESPSRGPF
      550      560      570      580      590      600

```

~~XXXXXXXXXX~~

This entry is from:  
SWALL (SPTR)

SWALL

Link



Print Entry

General Description References Comments Links Keywords Features Sequence

### General information

Entry name ESR1\_ORYLA  
Accession number P50241  
Created Rel. 34, 1-OCT-1996  
Sequence update Rel. 37, 15-DEC-1998  
Annotation update Rel. 40, 16-OCT-2001

01-10-1996

1-5

5

### Description and origin of the Protein

Description ESTROGEN RECEPTOR (ER) (ESTRADIOL RECEPTOR) (ER-ALPHA).  
Gene name(s) ESR OR NR3A1 OR MER.  
Organism source Oryzias latipes (Medaka fish).  
Taxonomy Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Actinopterygii; Teleostei; Euteleostei; Neoteleostei; Acanthomorpha; Acanthopterygii; Percomorpha; Atherinomorpha; Belontiiformes; Adrianichthyidae; Oryziinae;  
NCBI TaxID 8090

### References

- [1] Okada, H., Kawahara, T., Yamashita, I., RL Submitted (APR-1998) to the EMBL/GenBank/DDBJ databases.  
Position RP SEQUENCE FROM N.A.  
Comments RC STRAIN=D-RR; TISSUE=LIVER;
- [2] Kawahara, T., Yamashita, I., RT "Oryzias latipes genomic DNA for estrogen receptor. RL Submitted to the EMBL/GenBank/DDBJ databases.  
Position RP SEQUENCE FROM N.A.

### Comments

FUNCTION THE STEROID HORMONES AND THEIR RECEPTORS ARE INVOLVED IN THE REGULATION OF EUKARYOTIC GENE EXPRESSION AND A CELLULAR PROLIFERATION AND DIFFER IN TARGET TISSUES.

SUBUNIT BINDS DNA AS A HOMODIMER. CAN FORM HETERODIMER WITH ER- BETA (BY SIMILARITY).

SUBCELLULAR LOCATION NUCLEAR.

DOMAIN COMPOSED OF THREE DOMAINS: A MODULATORY N-TERMINAL DOMAIN, A DNA-BINDING DOMAIN, AND A C-TERMINAL STEROID-BINDING DOMAIN.

SIMILARITY BELONGS TO THE NUCLEAR HORMONE RECEPTOR FAMILY. NR3 SUBFAMILY.

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#### Database cross-references

EMBL	D28954;BAA25900.1;-
	AB033491;BAA86925.1;-
HSSP	P03372;1HCP.
	IPR000536;Hormone_rec_lig.
InterPro	IPR001292;Oest_recep.
	IPR001723;Strdhormone_receptor.
	IPR001628;zf-C4.
	PF00104;hormone_rec;1.
Pfam	PF02159;Oest_recep;1.
	PF00105;zf-C4;1.
PRINTS	PR00398;STRDHORMONER.
	PR00047;STROIDFINGER.
SMART	SM00430;HOLI;1.
	SM00399;ZnF_C4;1.
PROSITE	PS00031;NUCLEAR_RECEPTOR;1.

#### Keywords

Receptor, Transcription regulation; DNA-binding; Nuclear protein; Zinc-finger; Steroid-binding;

#### Features

Key	Begin	End	Length	Description
DOMAIN	1	185	185	MODULATING.
DNA_BIND	186	251	66	NUCLEAR RECEPTOR-TYPE.
ZN_FING	186	206	21	C4-TYPE.
ZN_FING	222	246	25	C4-TYPE.
DOMAIN	252	314	63	HINGE.
DOMAIN	315	620	306	STEROID-BINDING.
DOMAIN	299	307	9	POLY-GLY.
DOMAIN	320	323	4	POLY-LEU.
DOMAIN	511	515	5	POLY-LEU.
DOMAIN	576	579	4	POLY-GLY.

#### Sequence information

Length: 620 aa, molecular weight: 67729 Da, CRC64 checksum: DDCBD18C2B2BA522

MSKRQSSVQI RQLFGPALRS RISPASSELE TLSPPRLSPR DPLGDMYPEE SRGSGGVA  
DFLEGTYDYA APNPATTPLY SQSSTGYISA PLETNGPPSE GSLQSLGSGP TSPLVFVP  
PRLSPFMHPP SHHYLETTST PVYRSSHQGA SREDQCGSRE DTCSLGELGA GAGAGGF  
KDTRFCAVCS DYASGYHYGV WSCGCKAFF KRSIQGHNDY MCPATNQCTI DRNRKRS  
CRLRKCYEVG MMKGGVRKDR IRILRRDKRR TGVGDGDKV KGQEHKTVHY DGRKRS  
GGGGGGGRLS VTSIPPEQVL LLLQGAEPPI LCSRQKLSRP YTEVTMMTLL TSMADKELV  
MIAWAKKLPG FLQLSLHDQV LLESSWLEV LMIGLIWRSI HCPGKLIFAQ DLILDRNEGD

CVEGMTEIFD MLLATASRFR VLKLPPEEFV CLKAIILLNS GAFSFCTGTM EPLHNSAAVQ  
SMLDTITDAL IHYISQSGYL AQEQARRQAA LLLLLSHIRH MSNKGMEHLY SMKCKNKVPI  
YDLLLEMLDA HRLHHPVRAP QSLSQVDRDP PSTSSGGGGI APGSISASRG RIESPSRGF  
APSVLQYGGG RPDCTPALQD 620

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General	Description	References	Comments	Links	Keywords	Features	Sequence
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SRS 6.1.3 | feedback



>>SWALL:ESR1\_ORYLA ESTROGEN RECEPTOR (ER) (ESTRAD (620 aa)  
 initn: 4219 initl: 4219 opt: 4219 Z-score: 3109.1 bits: 587.6 E(): 3.1e-165  
 Smith-Waterman score: 4219; 100.000% identity (100.000% ungapped) in 620 aa  
 overlap (75-1934:1-620)

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	270	300	330	360	390	420
EP0111	DFLEGTYDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGLQSLGSGPTSPLVFPVPS					
SWALL:	DFLEGTYDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGLQSLGSGPTSPLVFPVPS					
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	450	480	510	540	570	600
EP0111	PRLSPFMHPPSHHYLETTSTPVYRSSHQASREDQCGSREDTCSLGELGAGAGAGGFEMA					
SWALL:	PRLSPFMHPPSHHYLETTSTPVYRSSHQASREDQCGSREDTCSLGELGAGAGAGGFEMA					
	130	140	150	160	170	180

	630	660	690	720	750	780
EP0111	KDTRFCAVCSYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPATNQCTIDRNRKSCQA					
SWALL:	KDTRFCAVCSYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPATNQCTIDRNRKSCQA					
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	810	840	870	900	930	960
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BNSDOCID: <XP 2181486A 1 >

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Winn R., Marine Environmental Research, vol. 46 (1-5), p. 130 (1998)

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# Aryl Hydrocarbon Receptor is Required for Prevention of Blood Clotting and for the Development of Vasculature and Bone in the Embryos of Medaka Fish, *Oryzias latipes*

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Higashi-Hiroshima 739-8527, Japan

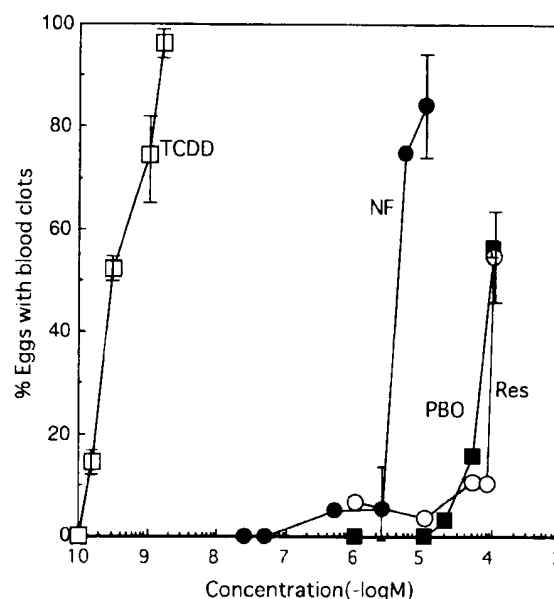
**ABSTRACT**—The aryl hydrocarbon receptor (AHR) is a member of ligand-activated transcription factors and conserved among vertebrates. To investigate the role of AHR in fish development, medaka embryos were treated with agonist (2,3,7,8-tetrachlorodibenzo-*p*-dioxin), antagonists ( $\alpha$ -naphthoflavone and resveratrol), and inhibitor (piperonyl butoxide) of cytochromes (Cyts) P450 encoded by a battery of target genes. These embryos were found to have similar abnormal phenotypes. Among the most consistent phenotypes were blood clotting and malformation of bone that were associated with vascular damages. These results thus indicate that control of AHR is important for proper development of fish embryos. AHR may control levels of Cyts P450 that are responsible for synthesis and metabolism of a toxic compound that caused the abnormal phenotypes. Complementary DNA fragments encoding AHR homologs were cloned from medaka embryos. AHR-specific mRNA was ubiquitously expressed in embryos and adult tissues.

**Key words:** aryl hydrocarbon receptor, blood clotting, bone formation, cytochrome P450, dioxin.

## INTRODUCTION

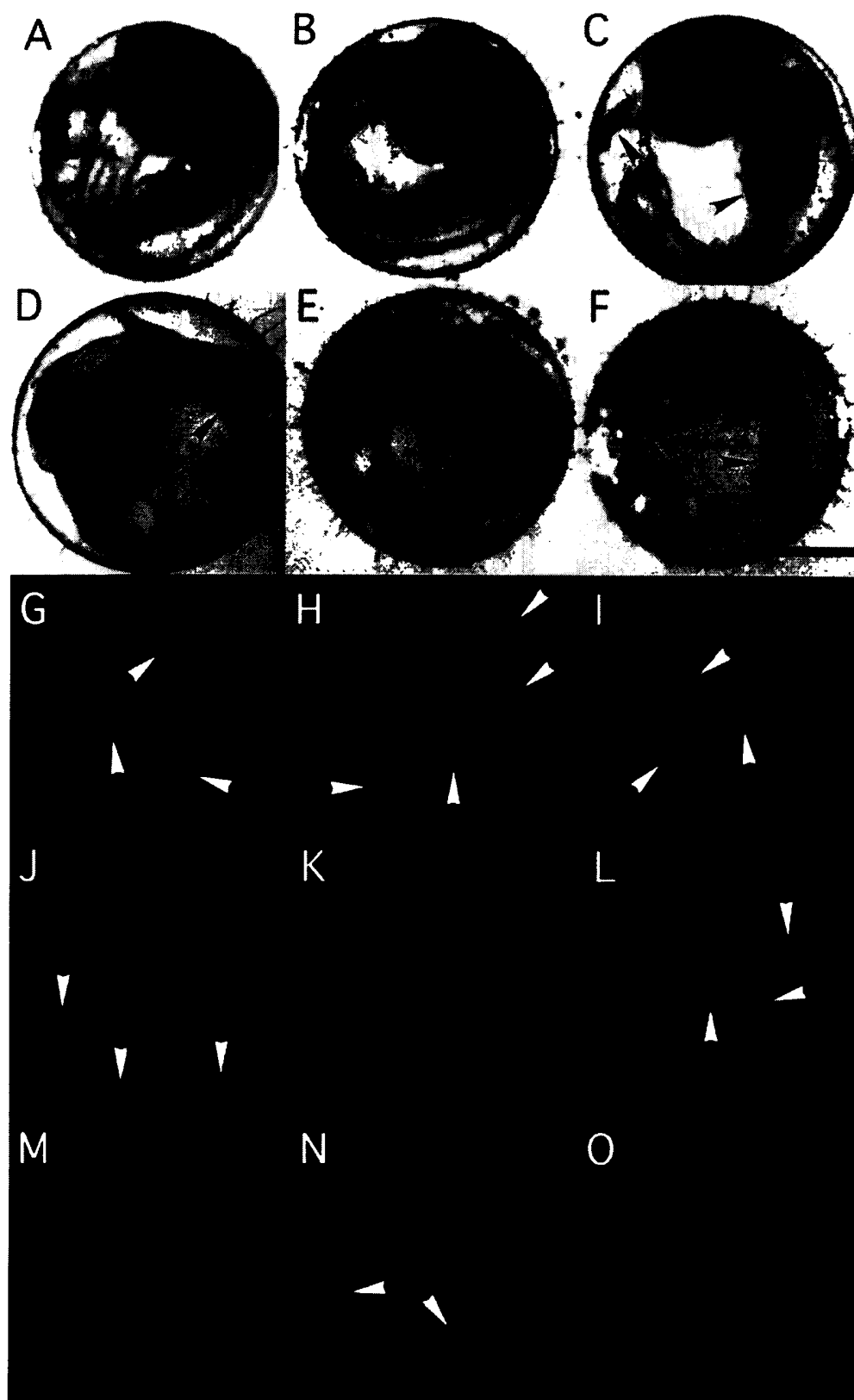
Planar halogenated hydrocarbons, such as 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD), are notorious environmental pollutants that are extremely toxic to early stages of vertebrate development (Peterson *et al.*, 1993). Hallmark signs of TCDD toxicity in fish sac fry are yolk sac edema, slowed blood flow, hemorrhage, and growth retardation culminating in mortality (Cantrell *et al.*, 1996; Henry *et al.*, 1997; Hornung *et al.*, 1999). Vascular damage, as assessed by TCDD-induced apoptotic cell death, is a key physiological mediator of the embryo toxicity (Cantrell *et al.*, 1996; Cantrell *et al.*, 1998). These chemicals bind to a ligand-dependent transcriptional factor called the aryl hydrocarbon receptor (AHR), resulting in the activation of a battery of genes encoding various cytochromes (Cyts) P450 that are responsible for degradation of the environmental contaminants (Hankinson, 1995; Guiney *et al.*, 1997; Guiney *et al.*, 2000). AHR is conserved among vertebrates, thus, may have arisen in an ancestral vertebrate as a detoxification system.

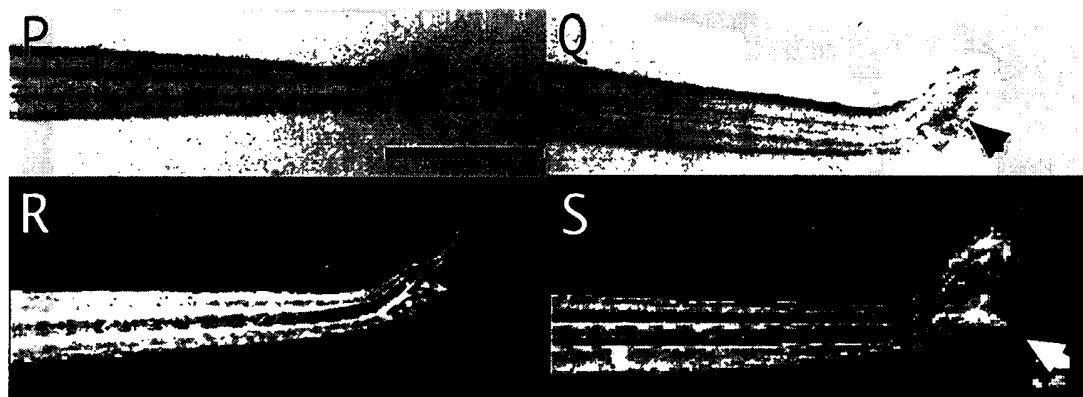
Although, to date, an endogenous ligand for AHR has not been found, AHR is ubiquitously expressed in most



**Fig. 1.** Effects of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD),  $\alpha$ -naphthoflavone (NF), resveratrol (Res), and piperonyl butoxide (PBO) on blood clotting during the embryo stage. Eggs were treated with TCDD, NF, Res, or PBO at the indicated concentrations until 6, 6, 4, or 5 dpf, respectively, and counted for blood clots.

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**Fig. 2.** Photographs of blood clots, yolk vein, and fin. Eggs and fry were treated with 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD),  $\alpha$ -naphthoflavone (NF), resveratrol (Res), or piperonyl butoxide (PBO) as follows and photographed for blood clots (A–F), yolk vein under green fluorescence (G–O), and fin (P–S): (A) mock-treated, 5 dpf; (B, C) 1.55 nM TCDD, 5 and 7 dpf; (D) 10  $\mu$ M NF, 5 dpf; (E) 100  $\mu$ M PBO, 5 dpf; (F) 100  $\mu$ M Res, 4 dpf; (G–I) mock-treated at 3, 5, and 7 dpf; (J, K) 1.55 nM TCDD, 5 and 7 dpf; (L, M) 10  $\mu$ M NF, 3 and 5 dpf; (N, O) 100  $\mu$ M PBO, 4 and 5 dpf; (P, R) mock-treated, 5-day post-hatching; and (Q, S) 0.155 nM TCDD, 5-day post-hatching. Arrows indicate blood clots (B–F, and Q), yolk veins (G–J, L, and N), and the constricted fin (S). Bar, 0.5 mm.

organs and cells in the body (Rowlands and Gustafsson, 1997). However, there is only a limited knowledge of developmental and physiological functions of AHR in the mouse (Gonzalez and Fernandez-Salguero, 1998), although the role of AHR in detoxification of environmental aryl hydrocarbons has been extensively studied *in vitro* (Hankinson, 1995). AHR-null mice were resistant to the acute toxicity (Fernandez-Salguero *et al.*, 1996) of and the teratogenic response (Mimura *et al.*, 1997) to TCDD, and found to have a number of abnormal phenotypes such as decreased accumulation of lymphocytes in the spleen and lymph nodes and reduction in liver size that are associated with accelerated rates of apoptosis (Fernandez-Salguero *et al.*, 1995), and difficulties in reproduction (Abbott *et al.*, 1999; Robles *et al.*, 2000). Thus, AHR is involved in the toxicity of and the teratogenesis by TCDD *in vivo*, and plays an important role in the development of the liver and the immune system, and in reproduction. However, no such function has been elucidated in other vertebrates.

Here we re-evaluated the role of AHR in chemical toxicity of TCDD in medaka fish embryos because there have been no pharmacological studies in fish using antagonist and also examined for any possible developmental and physiological function of AHR in medaka fish embryos using antagonists and Cyts P450 inhibitor. We found that AHR mediates TCDD toxicity such as blood clotting, malformation of bone, and regression of blood vessels, and that AHR is required for the embryonic development of vasculature and bone. To our knowledge, this is the first report of the developmental role of AHR in lower vertebrates.

## MATERIALS AND METHODS

### Fish and embryo culture

We used the d-rR strain of medaka fish, *O. latipes* (Kawahara and Yamashita, 2000). The fish were maintained at 25–26°C under

artificial photo-period of 14L 10D, and fed by powdered TetraMin (Tetra). Eggs were collected within 12 hr postfertilization (hpf), rinsed with tap water, and immersed in Yamamoto's salt solution (Yamamoto, 1969) with or without test chemicals. At least 30 eggs were used in each experiment. TCDD was purchased from Cambridge Isotope Laboratories, Inc. Antagonists,  $\alpha$ -naphthoflavone (NF) (Gasiewicz and Rucci, 1991; Merchant *et al.*, 1993) and resveratrol (Res) (Ciolino *et al.*, 1998; Casper *et al.*, 1999; Singh *et al.*, 2000), were from Sigma. Cyts P450 inhibitor, piperonyl butoxide (PBO) (Dahl and Hodgson, 1979; Testa and Jenner, 1981; Adams *et al.*, 1993), was from Tokyo Kasei Kogyo Co. These reagents were dissolved in acetone. The stock solutions were diluted over 1,000-fold with Yamamoto's solution and added to eggs of 12 hpf for NF, Res, and PBO or of 24 hpf for TCDD. The solvent was added to the mock-treated eggs as a control. The reducing agent, N-acetyl cysteine (NAC) (Sigma), was dissolved in Yamamoto's solution and added to 12 hpf eggs. Eggs and fry were cultured under the same condition as above (except without feed) and inspected for blood clotting under a dissecting microscope. Eggs and fry in which blood clots formed were counted.

Data are presented as mean  $\pm$  SEM. Statistical significance between values of control and experiment was assessed by Student's *t*-test.

### Observation of blood vessels

In order to observe the development of blood vessels, eggs were fixed with 4% paraformaldehyde for 3 days and observed under green fluorescence with a filter set (excitation filter, 546/10 nm, barrier filter, 590 nm) in Leica MZ FLIII stereo-fluorescence microscope. The fixed eggs were also dechorionated with forceps and stained with hematoxylin.

### Bone staining

In order to observe the bone development, calcified bone was stained with alizarin S essentially as described (Takeuchi, 1960). In brief, fish were anesthetized with 0.015% phenylurethane, skinned with forceps, treated with 2% KOH for 24 hr, and finally stained with 0.1% alizarin S solution. After washing in tap water, the fish were successively transferred to 50% and 70%, and finally embedded in 100% glycerin. Anesthetized fry were directly treated with 2% KOH for 2 h, fixed in 4% paraformaldehyde for 24 hr, then stained with alizarin S.

### Isolation of cDNAs encoding medaka AHR homologs

As PAS domain of AHR is highly conserved among vertebrates (Rowlands and Gustafsson, 1997), a corresponding region of cDNA was amplified with degenerated oligonucleotides (AhR-A1 and AhR-B1) as described (Hahn and Karchner, 1995) using total RNA from 6-day postfertilization (dpf) medaka embryos. The cDNA fragment was cloned in plasmid and sequenced. Based on the sequence, nested oligonucleotides were designed and 5' and 3' RACEs (rapid amplification of cDNA ends) were performed on the same RNA by using 5' and 3' RACE Systems (GIBCO BRL), yielding the remainder of the coding sequence, 5' and 3' untranslated regions, and polyadenylation sequence.

### RNA analysis

Total RNA was extracted from embryos and adult tissues as described (Kawahara *et al.*, 2000). RT-PCR (reverse transcription-polymerase chain reaction) analysis was done as described (Kawahara *et al.*, 2000) with the primers as follows for generation of the 437-bp cDNA encoding a part of PAS domain: poly(dT) oligonucleotide used for RT, and 5'-CCAGCAGGAGTTCAGGAGGA and 5'-ATTTACCCTTTGCGTCACA for PCR. Amplified DNA was electrophoresed in 1% agarose gel and stained with ethidium bromide.

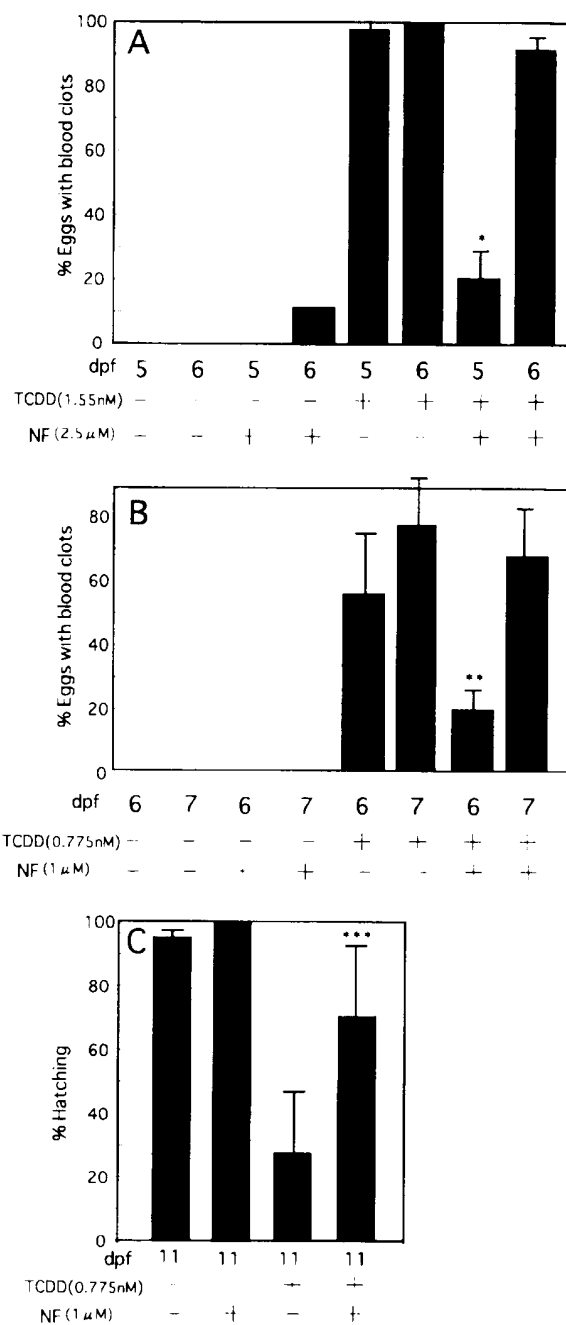
## RESULTS

### AHR mediates the toxic effects of TCDD on vascular development

We re-evaluated the toxic effects of TCDD on medaka embryos. To do this, embryos (1 dpf) were immersed in saline solution for medaka containing increasing concentrations of TCDD, and observed for any abnormal phenotype under a dissecting microscope (Fig. 1). Clearly visible signs of blood clotting were apparent after 4 days in caudal veins of TCDD-treated embryos (Fig. 2B), although blood cells were circulating in vasculature (Fig. 2J) but at a reduced rate. Blood clots were also found in yolk veins after 6 days (Fig. 2C), at that time, vascular structure was almost absent (Fig. 2K). In control embryos, yolk veins were apparent at 3 dpf (Fig. 2G) and developed progressively in a curve structure (Fig. 2A, H and I). Very small blood clots were occasionally found in yolk veins of normal embryos (less than 3%), but not scored in this study. These results are consistent with the previous observation that TCDD induces apoptosis of blood vessels (Cantrell *et al.*, 1996).

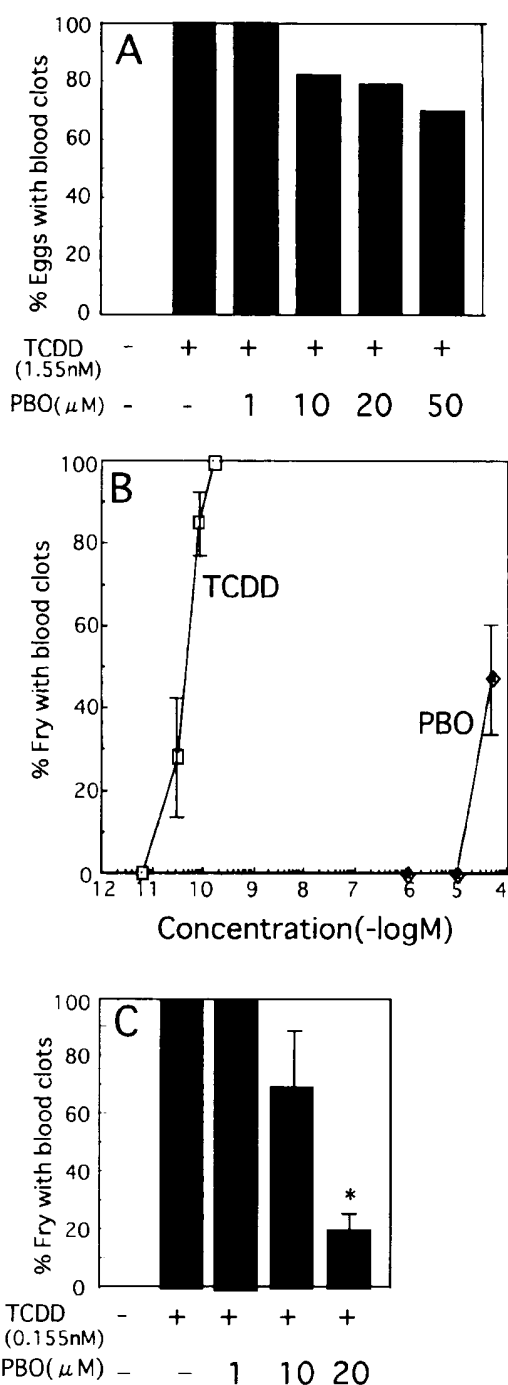
If TCDD induced the vascular damage through activation of AHR, the antagonist (NF) would reduce the extent to which blood clotting was detected. For this purpose, two different experiments were done, in which embryos were treated with high (1.55 nM) or medial (0.775 nM) concentration of TCDD (Fig. 3A or B, respectively). For both cases, addition of NF effectively suppressed blood clotting but only transiently (Fig. 3A and B). However, in the latter case, NF markedly enhanced the hatching success of TCDD-treated embryos, giving rise to almost complete hatching (Fig. 3C). These results indicate that TCDD-induced vascular damage is mediated through activation of AHR.

It is well known that TCDD-bound AHR activates transcription of a battery of genes encoding Cyts P450. If these



**Fig. 3.** Suppression by  $\alpha$ -naphthoflavone (NF) of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD)-induced blood clotting and mortality. **(A)** Eggs were treated with 1.55 nM TCDD and 2.5  $\mu$ M NF until 5 and 6 dpf as indicated, and examined for blood clotting. \* $P < 0.01$ . **(B)** Eggs were treated with 0.775 nM TCDD and 1  $\mu$ M NF until 6 and 7 dpf as indicated, and examined for blood clotting. \*\* $P < 0.2$ . **(C)** Eggs were treated as described in **(B)**, and examined for hatching rate at 11 dpf. \*\*\* $P < 0.05$ .

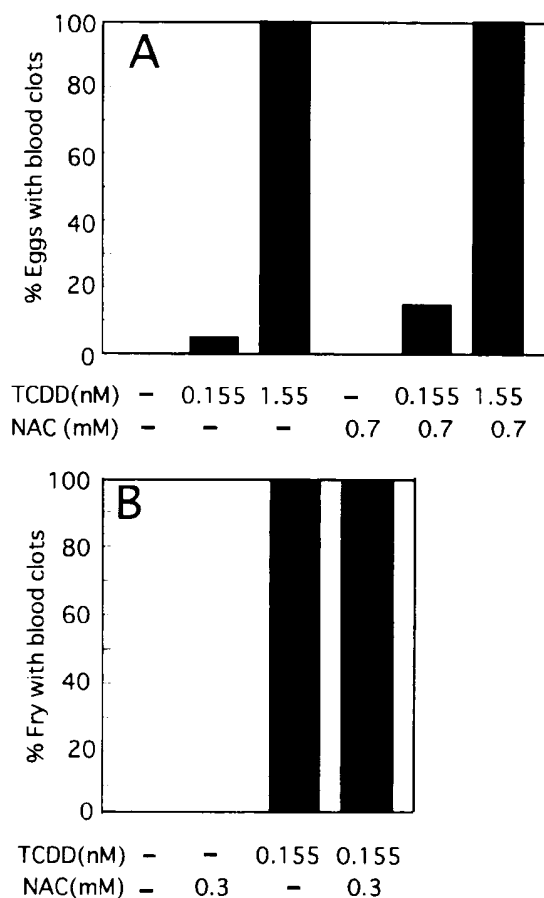
enzymes were involved in the TCDD-induced toxicity, an inhibitor of P450 would reduce the rate of TCDD-induced blood clotting. We therefore examined the ability of PBO to provide protection against high concentration (1.55 nM) of



**Fig. 4.** Suppression by piperonyl butoxide (PBO) of 2,3,7,8- tetra-chlorodibenzo-*p*-dioxin (TCDD)-induced blood clotting. (A) Eggs were treated with 1.55 nM TCDD and increasing concentrations (μM) of PBO as indicated until 5 dpf, and examined for blood clotting. (B) Eggs were treated with TCDD and PBO at the indicated concentrations until 5-day post-hatching, and examined for blood clotting in the caudal fin. (C) Eggs were treated with 0.155 nM TCDD and increasing concentrations of PBO as indicated until 5-day post-hatching, and examined for blood clotting in the caudal fin. \* $P < 0.05$ .

TCDD (Fig. 4A). Unexpectedly, PBO reduced the blood clotting rate only slightly; we cannot use higher concentrations of PBO because PBO itself induced blood clotting (described below). We therefore tried to seek for conditions under which lower concentrations of TCDD induce blood clotting effectively. We found that blood clots formed in the caudal fin (Fig. 2Q) after immersing embryos until 5-day post-hatching at subnanomolar concentrations of TCDD (Fig. 4B). Blood clots did not form in the control fin (Fig. 2P). Under the above condition, PBO effectively suppressed the adverse effect of TCDD (Fig. 4C). These results suggest that the TCDD-induced toxicity was caused by elevated expression of a certain Cyt P450.

Previous reports conclude that oxidative stress caused by TCDD-induced expression of Cyts P450 contributes to embryotoxicity and vascular damage associated with apoptosis, because the reducing agent, NAC, partially recovers



**Fig. 5.** N-acetyl cysteine (NAC) fails to suppress the 2,3,7,8- tetra-chlorodibenzo-*p*-dioxin (TCDD)-induced blood clotting. (A) Eggs were treated with TCDD (nM) and NAC (mM) at the indicated concentrations until 5 dpf, and examined for blood clotting. (B) Eggs were treated with 0.155 nM TCDD and 0.3 mM NAC as indicated until 5-day post-hatching, and examined for blood clotting in the caudal fin.



the TCDD-induced embryotoxicity (Cantrell *et al.*, 1996): they observed 41% survival of the embryos that had been treated with 28 nM TCDD for 2 hr and released in 0.1 mM NAC until 3 days posthatch, in contrast to 2% survival of the embryos that had been treated with TCDD and released in water. The ability of NAC to inhibit TCDD-induced toxicity was re-assessed by adding 0.7 mM (Fig. 5A) or 0.3 mM (Fig. 5B) NAC to eggs before and during the treatment with TCDD. NAC could not inhibit the blood clotting induced by 0.155 or 1.55 nM TCDD. NAC itself induced blood clotting at more than 0.9 mM (data not shown). These results suggest that general oxidative stress is not responsible for the TCDD-induced blood clotting.

#### Vascular damage induced by antagonists (NF and Res) and Cyts P450 inhibitor (PBO)

At the initial experiments determining the concentrations of reagents used, we found that NF, Res, and PBO induced blood clotting at higher concentrations than those used for suppression of TCDD-induced toxicity (Fig. 1). Blood clots formed in caudal and yolk veins (Fig. 2D-F). Yolk veins developed normally at the early time of incubation (up to 4 dpf) (Fig. 2L and N), but their regression was apparent at the time when blood clots formed in yolk veins (at 5 dpf) (Fig. 2M and O). These results suggest that either inactivation of AHR by NF and Res or inhibition of certain Cyts P450 by PBO caused vascular damage and blood clotting.

If the hypothesis were true, antagonist of AHR and Cyts P450 inhibitor would act synergistically to cause toxicity. We examined the synergy between low concentrations of NF (2.5  $\mu$ M) and PBO (20  $\mu$ M) that alone did not show any effect. Combination of these chemicals clearly increased the

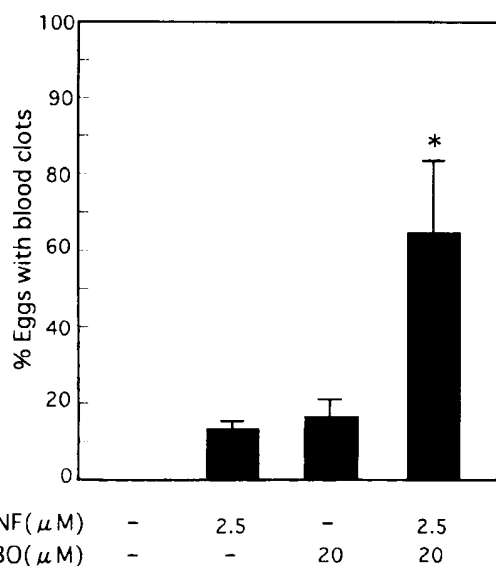


Fig. 6. Synergistic effects of  $\alpha$ -naphthoflavone (NF) and piperonyl butoxide (PBO) on blood clotting. Eggs were treated with NF and PBO at the indicated concentrations ( $\mu$ M) until 6 dpf, and examined for blood clotting. \* $P < 0.2$ .

rate of blood clotting (Fig. 6). We therefore conclude that control of AHR activity and levels of Cyts P450 is required for proper development of vasculature in fish.

#### Malformation or degeneration of bone induced by TCDD, NF, and PBO

During the experiments by incubating eggs with lower concentrations of TCDD (less than 80 pM) until 7 dpf, most eggs developed normally in appearance and blood clots did not form. The eggs were transferred to Yamamoto's solution, then to aquaria after hatching, and reared to adult by normal diet as usual. Unexpectedly we found that these fish were deformed in shape like wavy mutants (Takeuchi, 1960). We examined the bone development by staining with alizarin S. The vertebral column of TCDD-treated fish curved dorso-ventrally and laterally (Fig. 7A and B). Neural and haemal spines were short in length and deformed (Fig. 7B). NF also suppressed the TCDD-induced toxicity on bone formation (Fig. 7C), indicating the involvement of AHR.

We examined the effect of TCDD on the embryonic bone formation by incubating eggs with TCDD until 5 days post-hatching. The staining of the fry with alizarin revealed the absence of calcification in the posterior region of spinal cord and in spines (Fig. 7D and E). We also found that caudal fins were round in shape and constricted (indicated by arrow in Fig. 2S) in the TCDD-treated fry.

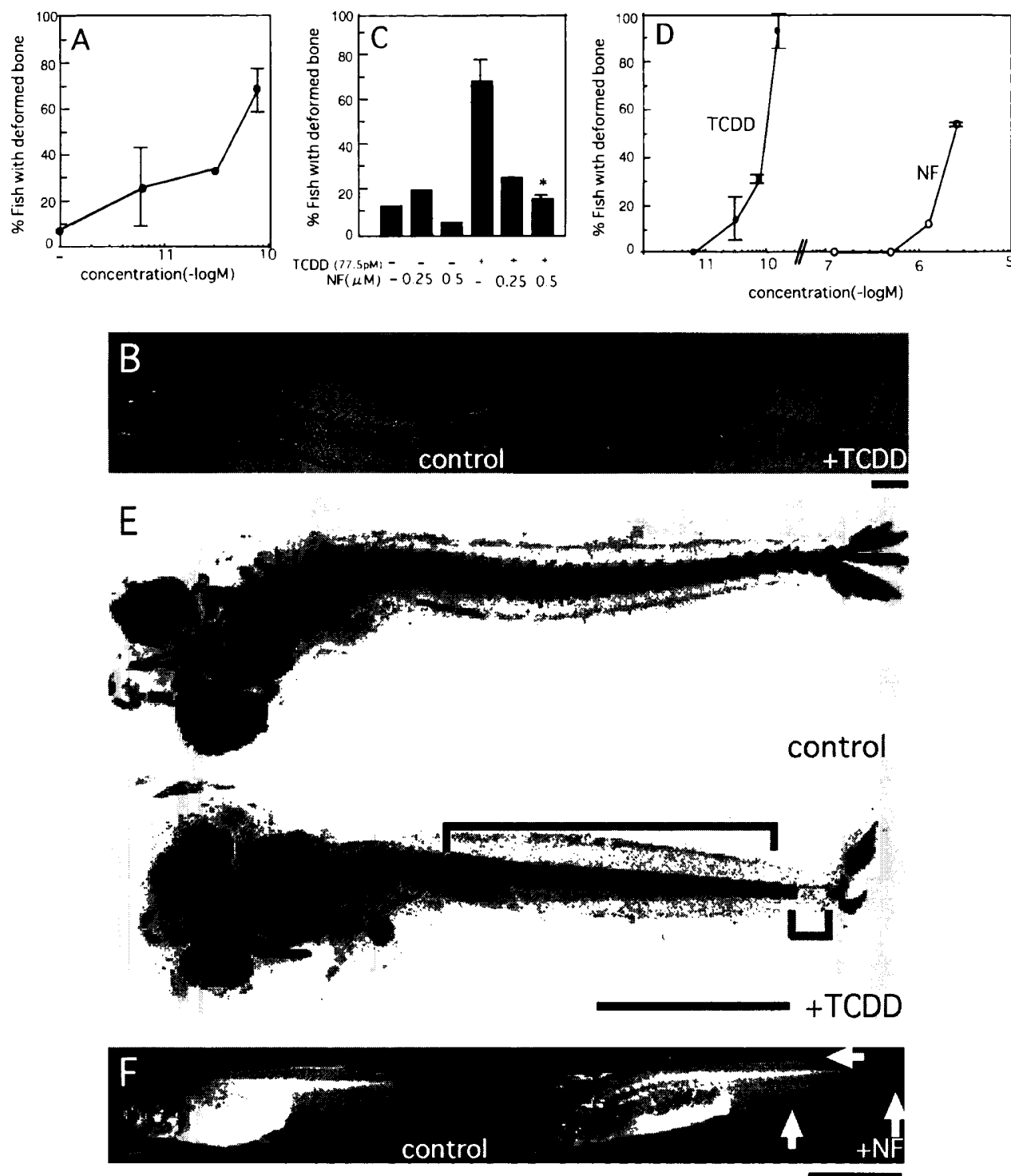
In order to examine the possible function of AHR and Cyts P450 in the embryonic bone formation, eggs were treated with NF or PBO until 5 days post-hatching. The treatment with NF (2.5  $\mu$ M) did not cause blood clotting in any portion of the fry (data not shown), which was different from the result with TCDD (Figs. 2Q and 4B). However, the treatment also caused degeneration of the posterior end of the spinal cord, but with normal development of spines (Fig. 7D, data not shown). PBO (50  $\mu$ M) also caused the same defect in bone formation as that NF did (data not shown).

We further examined whether NF affects homeostasis of adult fish. To do this, adult fish which had been reared by normal diet for 2 months were fed by NF-containing diet (2 mg NF/g diet) for 2 months. During the cultivation, population of fish lacking posterior fins including anal, caudal, and dorsal fins appeared after a month and became increasing near to 100% by two months (Fig. 7F).

Taken together, these results suggest that hyperactivation of AHR by TCDD is toxic to the embryonic development of bone and caudal fin, that AHR is required for proper development of bone and homeostasis of posterior fins, and that a certain Cyt P450 is also required for bone development.

#### Isolation and characterization of cDNAs encoding AHR homologs of medaka fish, and ubiquitous expression of AHR mRNA

We first obtained four independent cDNA clones (clones 1, 2, 3 and 4) corresponding to PAS domain (Fig. 8A). These clones were found to be identical by sequencing.

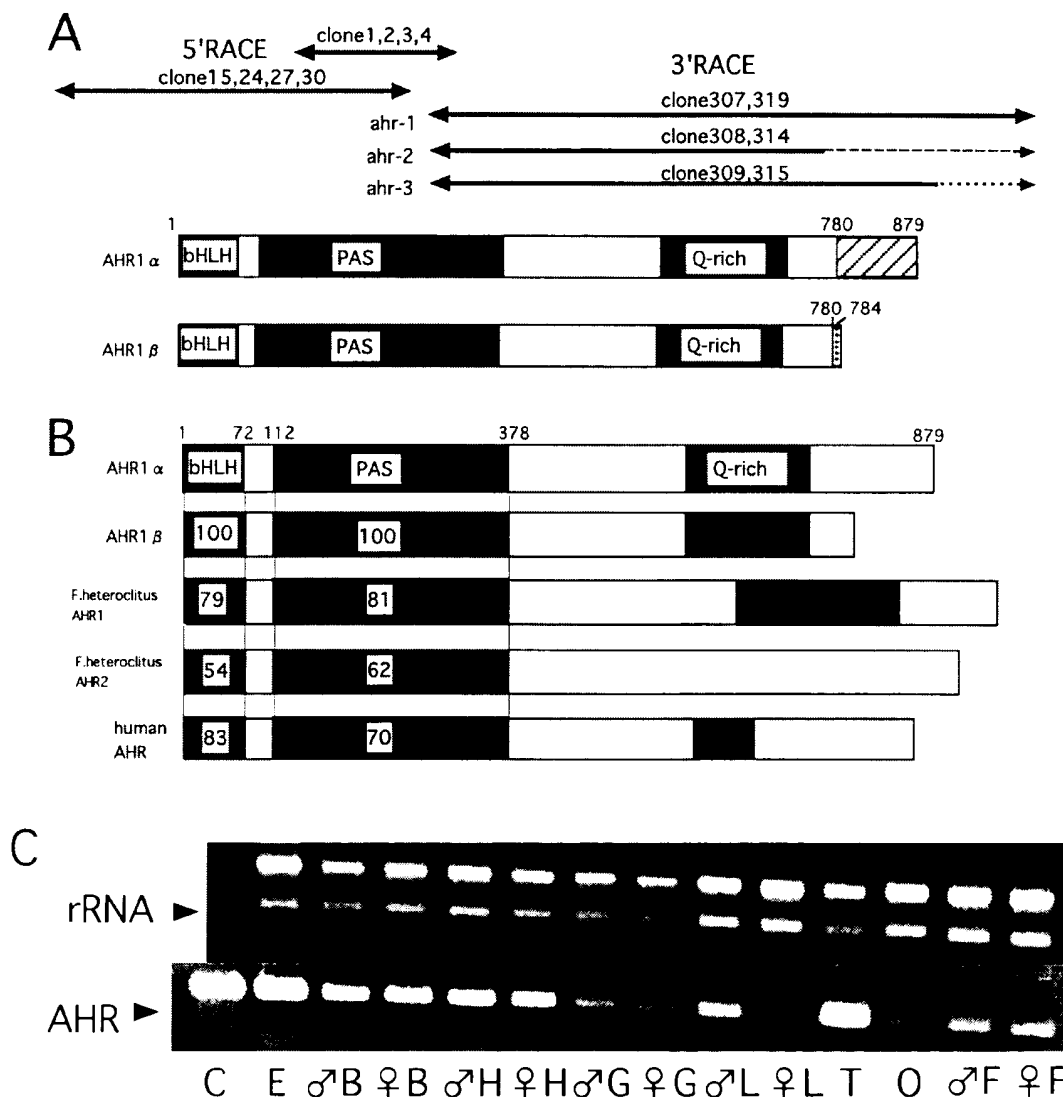


**Fig. 7.** Effects of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD) and  $\alpha$ -naphthoflavone (NF) on bone formation. (A) Eggs were treated with TCDD at the indicated concentrations until 7 dpf, and reared to adult under TCDD-free condition. The adult fish were examined for bone formation after staining with alizarin. (B) Alizarin-stained bone of mock-treated (control) and TCDD (77.5 pM)-treated fish in (A). (C) Eggs were treated with 77.5 pM TCDD and NF at the indicated concentrations ( $\mu$ M) until 7 dpf, reared to adult under normal condition, and examined for bone formation. \* $P < 0.05$ . (D) Eggs were treated with increasing concentrations of TCDD and NF until 5-day post-hatching, and examined for bone formation. (E) Alizarin-stained bone of mock- (control) and TCDD (0.155 nM)-treated fish in (D). Spines and posterior spinal bone are absent in the TCDD-treated fry as noted. (F) Normal adult fish were fed by NF-containing diet (2 mg NF/g diet) for 2 months, and photographed. Arrows indicate the degenerated fins. Bar, 1 mm in (B) and (E), and 5 mm in (F).

Next, 5' and 3' RACEs were performed, yielding four (clones 15, 24, 27 and 30) and six (clones 307–309, 314, 315, and 319) independent clones, respectively (Fig. 8A). Four clones from 5' RACE were identical. Six clones from 3' RACE were subdivided into three identical pairs, which differ from each other only in the 3' proximal sequences denoted by broken and dotted lines in Fig. 8A. Thus, we obtained three different cDNAs, named *ahr-1*, -2, and -3 (DDBJ accession numbers AB065092, AB065093, and AB065094, respectively). However, *ahr-1* and *ahr-3* encoded the same protein (AHR1 $\alpha$ ), and *ahr-2* encoded another homolog (AHR1 $\beta$ ). AHR1 $\alpha$  and

AHR1 $\beta$  differ from each other in the C-terminal peptides (amino acid 780–879 and 780–784) denoted by shaded and dotted boxes (Fig. 8A).

AHR1 $\alpha$  and AHR1 $\beta$  are composed of 879 and 784 amino acids with calculated molecular weights of 95.5 and 85.3 kDa, respectively. Both proteins may be classified into a type of AHR1 because they are most homologous to AHR1 of the teleost *Fundulus heteroclitus* (Karchner *et al.*, 1999) (Fig. 8B). The medaka AHR1 $\alpha$  and AHR1 $\beta$  are also composed of three conserved domains such as basic-helix-loop-helix (bHLH), Per-ARNT-Sim (PAS), and glutamine-rich



**Fig. 8.** Schematic drawing of the cDNAs cloned and the deduced proteins, and ubiquitous expression of AHR mRNA. **(A)** Inserts in the plasmid clones are shown on the deduced proteins (AHR1 $\alpha$  and AHR1 $\beta$ ). Plasmid numbers are marked on the corresponding inserts. The three cDNAs which differ from each other only in the 3' terminal sequences (denoted by broken and dotted lines) are named *ahr-1*, -2, and -3. AHR1 $\alpha$  and AHR1 $\beta$ , in which three conserved domains are marked by bHLH, PAS, and Q-rich, differ from each other only in the C-terminal short peptides marked by shaded and dotted boxes. **(B)** Identity (%) of amino acid sequence among bHLH and PAS domains of AHRs from medaka, *F. heteroclitus* (killifish), and human (Dolwick *et al.*, 1993). **(C)** RT-PCR analysis of total RNAs from medaka embryos (6 dpf) and adult tissues. Symbols: B, brain; C, the control band amplified from the cDNA; E, embryo; F, caudal fin; G, gill; H, heart; L, liver; O, ovary; and T, testis. Ribosomal RNAs in the RNA samples are also shown.

(Q) domains (Rowlands and Gustafsson, 1997) (Fig. 8B).

Expression of AHR mRNA was analyzed by RT-PCR on total RNAs prepared from medaka embryos and adult tissues such as brain, fin, gill, heart, liver, ovary, and testis. AHR mRNA was detected in all samples tested, and in large amounts in embryos and testis (Fig. 8C).

## DISCUSSION

### TCDD-induced vascular and bone damages through hyperactivation of AHR

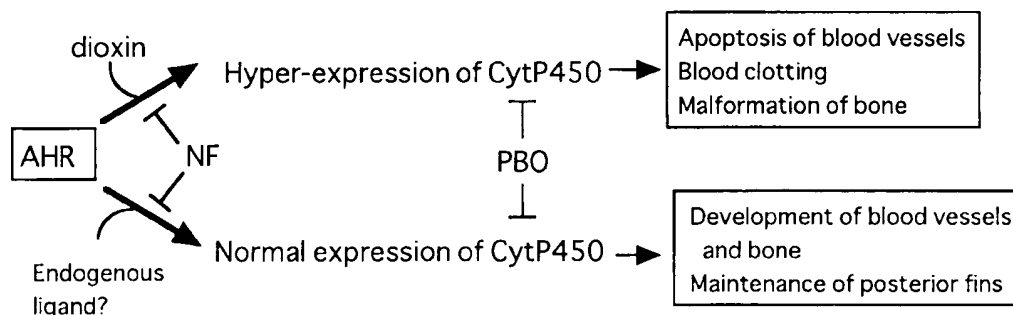
TCDD is the most potent toxicant for vertebrate species. Exposure of vertebrate embryos to TCDD can result in various acute and chronic toxicities such as reproductive failure, teratogenic abnormalities, and immunological dysfunction (Peterson *et al.*, 1993). In fish, vascular damage is the most pronounced adverse effects of TCDD exposure during embryonic development. Vascular hemorrhaging, regression of blood vessels, pericardial sac edema, and reduced circulation are hallmark indicators that vascular function is compromised in the developing embryos (Cantrell *et al.*, 1996; Henry *et al.*, 1997; Hornung *et al.*, 1999; Guiney *et al.*, 2000). The vascular lesions have been demonstrated to be associated with apoptosis and induced expression of Cyt P450 1A in blood vessels of medaka embryos (Cantrell *et al.*, 1998). In the present study, we re-examined the TCDD-induced vascular damage in medaka embryos by observing blood clotting and regression of blood vessels. We found that these vascular damages can be suppressed, but transiently, by antagonist, NF (Fig. 3), giving a convincing evidence that the TCDD-induced vascular damage is mediated through hyperactivation of AHR. The transient suppression may be explained by the fact that TCDD, but not NF, is very stable *in vivo* against catabolic activities of Cyts P450 (Miniero *et al.*, 2001). Although the damage can also be suppressed by Cyts P450 inhibitor, PBO (Fig. 4C), general oxidative stress caused by Cyts P450-mediated oxidative reactions may not be responsible for the

TCDD-induced damage, in inconsistent with the previous conclusion (Cantrell *et al.*, 1996), because reducing agent, NAC, could not recover the damage in vasculature (Fig. 5) or also in bone (data not shown). We assume that a toxic compound that may be accumulated *in vivo* by elevated levels of Cyt P450 is responsible for the TCDD-induced pathology (Fig. 9).

We also found that embryonic treatment with picomolar concentrations of TCDD causes malformation of bone in adult fish (Fig. 7). The treatment did not give apparent complications including blood clotting in the hatching fry, thus, the bone staining is the most sensitive method for detecting TCDD toxicity. The bone deformity could also be recovered by co-treatment with the antagonist (Fig. 7C), implying the role of hyperactivated AHR. TCDD may directly act on bone, because it inhibits osteogenesis in bone-forming cultures of chicken and rat cells (Gierthy *et al.*, 1994; Singh *et al.*, 2000). Treatment of medaka fish with TCDD from the egg stage to post-hatching also caused developmental defects in bone formation at the posterior region of vertebral column and at spines (Figs. 7D, E). However, it may be possible that these defects occurred secondarily to vascular damage, because blood clots formed at the base of the caudal fin under the same condition (Fig. 2Q).

### AHR is required for prevention of blood clotting and for proper development of vasculature and bone in medaka fish

AHR is conserved among vertebrates, and ubiquitously expressed in embryos and adult tissues. In the present study, we have cloned three different cDNAs encoding two AHR homologs from medaka fish, *O. latipes* (Fig. 8). The two homologs obtained may belong to a type of AHR1 by amino acid sequence similarity, thus named AHR1 $\alpha$  and AHR1 $\beta$ . They differ from each other only in C-terminal short peptide, and may be derived from alternative splicing. AHR1 mRNA was also ubiquitously expressed in medaka embryos and adult tissues, suggesting developmental and physiolog-



**Fig. 9.** Model for the role of AHR in the TCDD (dioxin)-induced toxicity, the development of blood vessels and bone, and the maintenance of posterior fins in the medaka fish, *O. latipes*. TCDD-bound AHR induces hyper-expression of a certain Cyt P450, resulting in the toxicities such as apoptosis of blood vessels, blood clotting, and malformation of bone. Either the antagonist (NF) or the Cyts P450 inhibitor (PBO) can suppress the TCDD-induced toxicity. An endogenous ligand is bound to and constitutively activates AHR. The activated AHR is responsible for normal expression of a certain Cyt P450 that is required for the development of blood vessels and bone and homeostasis of posterior fins. *In vivo* inhibition of AHR and Cyt P450 by NF and PBO, respectively, causes developmental abnormalities in vasculature and bone.

ical roles in medaka fish.

To investigate the role of AHR in fish development and physiological homeostasis, medaka embryos (12 hpf) were treated with the antagonists, NF and Res. These compounds did not cause any apparent defects until 4 dpf, but displayed developmental toxicities such as blood clotting and regression of blood vessels at 5 dpf (Figs. 1 and 2). Blood clotting may be caused by regression of blood vessels, because platelet adhesion to subendothelial collagens and activation by components of the extracellular matrix are crucial for blood coagulation (Nieswandt *et al.*, 2001). NF also caused the malformation of bone at 5-day post-hatching (Fig. 7D) and the regression of posterior fins such as anal, caudal, and dorsal fins at the adult period (Fig. 7F). These results suggest the presence of an endogenous ligand for AHR and that constitutive activation of AHR is specifically required for the development of blood vessels and bone and for the maintenance of posterior fins (Fig. 9).

Ligand-bound AHR activates transcription of a battery of genes including various Cys P450. If levels of a certain Cyt P450 were controlled by AHR bound to an endogenous ligand and required for proper development of blood vessels and bone, the well-known inhibitor (PBO) of the enzymatic activity of Cys P450 would induce the same developmental defect as did the antagonist. Treatment of embryos with PBO specifically induced blood clotting, regression of blood vessels (Figs. 1 and 2), and degeneration of the posterior end of spinal cord (data not shown) at the same developmental stage as did the antagonist, suggesting the importance of Cyt P450, the identity of which is, however, unknown (Fig. 9). The synergistic effects exerted by NF and PBO (Fig. 6) also support the hypothesis. We assume that a certain Cyt P450 is responsible for degradation (or catabolism) of a toxic compound that caused the developmental abnormalities.

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Tokio Yamamoto In: "Medaka, Biology and Strains" (T. Yamamoto, ed.), Yugakusya Publ. (1975), pp. 17-29.

## **Systematics and Zoogeography**

The killifishes, or Cyprinodontiforms are small fresh and brackish water fishes of worldwide distribution in tropical and temperate latitudes.

### **Previous classification of the order Cyprinodontes**

The classification of the order Cyprinodontes Agassiz (equivalent to Microcyprini Regan) has been worked out by Gill (1865, 1874), Regan (1909, 1911), Hubbs (1924, 1926) and Myers (1931, 1938). The classification followed here is mostly according to Hubbs and Myers and is cited from Kulkarni (1940) who erected a new family Horaichthyidae represented by a remarkable Indian henpecked killifish, *Horaichthys setnai*. However, substituting for the terms Amblyopsoidea and Poecilioidea, the suborders Amblyopsoidei and Cyprinodontoidei are used here, respectively. The subfamily Tomeurinae is removed from the family Poeciliidae to erect a new family Tomeuridae as suggested by Hubbs in his letter to S. L. Hora (India) in 1938. Representative genera are given in parentheses following family names.

Order Cyprinodontes (Microcyprini)

Suborder Amblyopsoidei

Family Amblyopsidae (*Chologaster*, *Amblyopsis*)

Suborder Cyprinodontoidei

Family Cyprinodontidae (*Cyprinodon*, *Fundulus*,

*Aplocheilus*, *Panchax*, *Oryzias*)

Family Goodeidae (*Goodea*)

Family Poeciliidae (*Poecilia*, *Gambusia*, *Xiphophorus*)

Family Jenynsiidae (*Jenynsia*)

Family Anablepidae (*Anableps*)

Family Tomeuridae (*Tomeurus*)

Family Adrianichthyidae (*Adrianichthys*, *Xenopoecilus*)

Family Phallostethidae (*Phallostethus*, *Gulaphallus*)

Family Horaichthyidae (*Horaichthys*)

The classification listed here has been generally held by ichthyologists until 1962.

As to the status of *Oryzias*, Myers (1931) considered it to represent a monogeneric tribe of the subfamily Fundulinae. Later (1956), he revised his earlier classification, and considered *Oryzias* to represent a monogeneric subfamily of the Cyprinodontidae, the Oryziatinae.

## Classification of new order Atherinoformes

Rosen (1962) presented evidence which indicates a relationship of the Amblyopsidae (North American cave fishes) with the percopsiform genera and, more distantly with the gadiforms. He isolated the cave fishes as a new order, the Amblyopsiformes, and recommended its alignment near the Percopsiformes and Gadiformes in a phyletic sequence.

In 1964, Rosen has made drastic taxonomic re-arrangements of the halfbeaks, killifishes, silversides, and their relatives. The outset of his re-arrangements was osteological analyses of the adrianichthyid fishes of Celebes, which were found to have a mixture of beloniform, cyprinodontiform, and mugilform features. Then, his investigation was broadened to include representatives of all these groups as well as a species of phallostethid.

In consequence of reasonable osteological diagnoses, he erected a new order Atherinoformes which includes the excoetoids, scomberesocoids. On the basis of osteological evidence, he separated the medaka (*Oryzias*) from cyprinodontoids, placed it in adrianichthyoids and erected a new family Oryziatidae.

To visualize Rosen's account on osteological difference between cyprinodontoids and adrianichthyoids, the presentation of the schema of the skull of the generalized teleosts as shown in Fig. 2-1 may be apropos.

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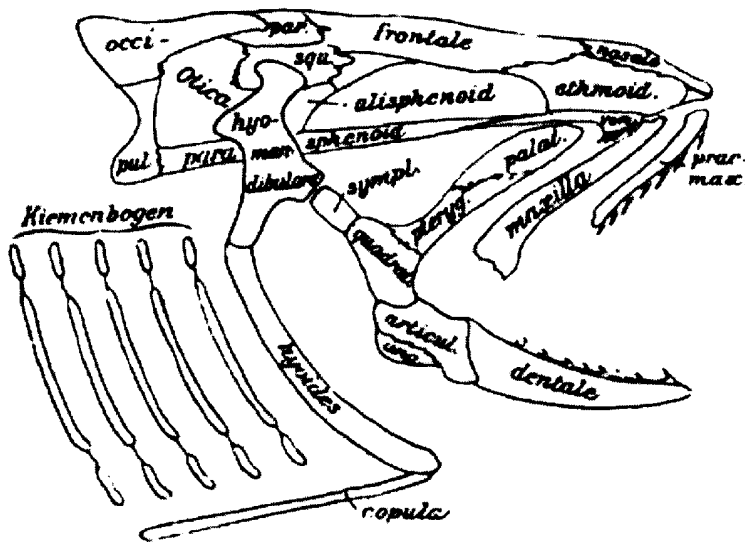


Fig. 2-1. A diagram of teleostean skull. Opercula and Infraorbitalia are removed. ang.=angular, articul.=articular, occi.=occipital, palat.=palatine, p.=quadrate, squ.=squamosa, sympl.=symplectic, vom.=vomer.

After R. Goldschmidt' E. Selenkas Zoologisches Taschenbuch fur Studierende. 1912 Leipzig, George Thieme.

In cyprinodontoid killifishes, bones of the jaws and the palatoquadrate arch are in such a construction that the premaxilla is protractile. In adrianichthyoid killifishes, on the other hand, the premaxillae are not protractile. The Adrianichthyidae are fishes of small size confined to the fresh-water lakes of Celebes. Two species, *Adrianichthys kruyti* and *Xenopoecilus sarasinorum*, are known. *Xenopoecilus* is characterized by having a large horse-shoe shaped mouth, an enormous ethmoideum and a single, median supraoccipital process formed by fusion of embryologically paired elements; "a cup-like excavation on the distal tip of the autopalatine that is capped by a large ball of cartilage and a discoidal sesamoid bone; a dorsal enlargement of the palatopterygoid arch with a prefrontal (Fig. 2-2); a maxilla that is carried on the upper edge rather than on the outer face of the posterior end of the premaxilla; a premaxilla that lacks a hooked or pointed posteroventral process; a tremendously reduced articular bone without a coronoid process that is almost wholly contained within the posterior part of the dentary; the articulation of the first pleural rib on the third rather than on the second vertebra; pelvic girdles that are not in contact medially and that have a long lateral spur extending upward between ribs; a dorsoventrally asymmetrical caudal skeleton with one or two very slender, rod-like epurals, and a caudal fin that is divided into indistinct upper and lower lobes by having a large gap between rays that articulate with the upper and lower hypural plates on the terminal half-centrum. (Rosen, 1964)

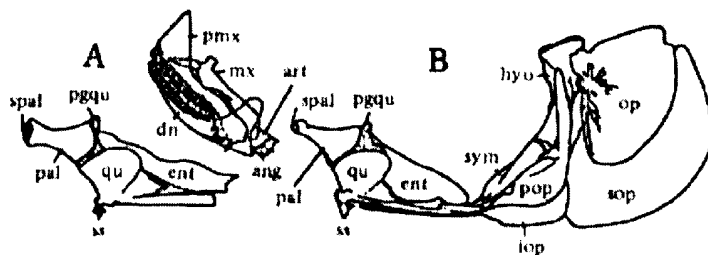


Fig. 2-2. Jaws and jaw suspension in adrianichthyoid killifishes. A. Jaws and palatopterygoid arch in *Oryzias latipes* (Temminck and Schlegel). b. Jaw suspension and opercular apparatus in *Xenopoecillus sarasinorum* (Popta). Note sesamoid bone below quadrate and bony cap over tip of palatine in A. and B. Note in A that lower arm of premaxilla lies over maxilla, large coronoid process on dentary, and absence of similar coronoid elevation on articular. Ang = angular, art = articular, dn = dentary, ent = entopterygoid (mesopterygoid), hyo = hyomandibular, iop = interoperculum, mx = maxilla, op = operculum, pal = palatine (autopalatine with or without dermopalatine), pgqu = pterygoquadrate cartilage, pmx = sesamoid bone capping autopalatine, ss = sesamoid bone, sym = symplectic. Rosen, 1964.

Rosen pointed out that except for the enlarged jaws and the presence of a median supraoccipital process, all the above features described within quotation marks can be identified in *Oryzias* (Fig. 2-2) but in no other killifishes so far as known.

It is therefore apparent that adrianichthyids and the medaka are intimately related and that they constitute a distinct subgroup of the killifishes, the adrianichthyoids, containing the families Adrianichthyidae (*Adrianichthys* and *Xenopoecilus*), Oryziatidae (*Oryzias*), and Horaichthyidae (*Horaichthys*), in contrast to the remainder of the families which are grouped together as cyprinodontoids (Cyprinodontoidea). Basing on Rosen's (1964) findings, Turner (1965) conveniently enumerated difference between cyprinodontoids and *Oryzias* as follows:

Cyprinodontoidae	<i>Oryzias</i>
1. First pleural rib on second vertebra.	First pleural rib on third vertebra.
2. Pelvic girdle bones joined mid-ventrally; no upright lateral spur.	Pelvic girdle bones not joined mid-ventrally; an upright lateral spur present.
3. Lower end of premaxilla bone expanded or hooked and sandwiched between the lower end of maxilla bone and dentary bone (lower jaw).	Lower end of premaxilla bone not expanded, and dorsal to the maxilla bone rather than between it and the dentary bone.
4. Hypural plates often fused.	Hypural plates never fused.
5. Hypochordal musculature entirely absent.	Hypochordal musculature present.
6. Caudal fin never incipiently lobed.	Caudal fin incipiently lobed.

The family Horaichthyidae erected by Kulkarni (1940) comprises a single species, *Horaichthys setnai*. It is a small translucent oviparous fish inhabiting brackish waters and estuaries in the province of Bombay, India. Osteological study (Kulkarni 1948) showed that its head skeleton is closely allied to that of *Oryzias* but greatly different from that of *Aplocheilichthys*. *Horaichthys*, however, is different from known species of *Oryzias* in having a larger number of the anal fin-rays (about 28 to 32).

In the male, six anterior rays of the anal fin are separated from the rest of the fin and modified into an elaborate male organ (gonopodium). Of six rays the third, fourth and fifth ones are profoundly modified forming the 3-4-5 complex. (Fig. 2-3). In the female right pelvic fin is usually absent. The genital opening of the female is situated on the left ventral side and is surrounded by genital pads. *Horaichthys* is supposed to have evolved from *Oryzias*, but as the development of the gonopodium in association with the henpecked sexual behavior is so remarkable that Kulkarni (1940) has proposed to erect a new family rank for this fish.

The male appears to be always afraid of the female which on occasions chases him away. At the time of mating, "the male swims below and behind her at a distance of about 2 to 3 cm. He then darts towards her on the left with almost lightning speed. As he approaches his mate he lashes out the gonopodium sideways almost at right angles to his body and strikes its terminal end against her genital opening. The spermatophores are transferred to the female in this momentary contact, and become attached by their distal hooks."

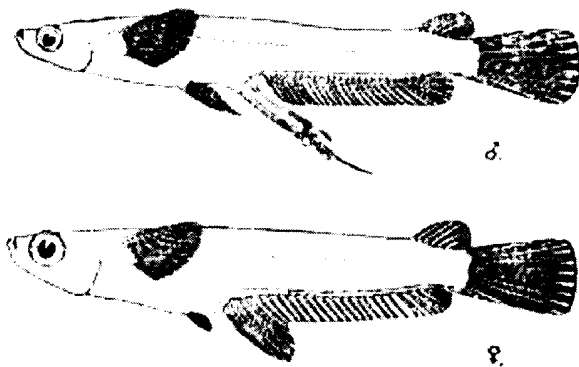


Fig.2-3. Lateral view of a male and a female specimen of *Horaichthys setnai*.  
x 4 Kulkalni, 1940.

A special feature of *Horaichthys* is that the testis produces special sperm capsules of spermatophores (2-300 in number) instead of ordinary semi-fluid milt with suspended sperms.

A spermatophore is a tiny hyaline body (0.6 mm long and 0.1 mm thick), the broad part of which contains mass of sperms. At the tapering end, there is a pointed cap with stiff hooks and barb-like structures which point backwards. It is with the aid of these hooks and barbs that the spermatophore get attached near the genital opening of the female.

There is no permanent opening on the spermatophore for the liberation of sperms. Before liberation of sperms, a small bulging appears at the neck of the tapering spermatophore and begins to enlarge. When the protuberance becomes sufficiently large, an opening is formed at its tip by rupture of membrane and sperms are liberated. They swim into the genital pore of the female.

The following is the classification of the new order Atheriniiformes by Rosen (1964), representative species being given in parentheses.

Suborder Exocoetoidei

Superfamily Exocoetoidea

Family Hemiramphidae (*Hemiramphus*)

Family Exocoetidae (*Exocoetus*)

Superfamily Scomberesocoidea

Family Belonidae (*Ablennus*)

Family Scomberesocidae (*Cololabis*)

Suborder Cyprinodontoides

Superfamily Adrianichthyoidea

Family Oryziatidae (*Oryzias*)

Family Adrianichthyidae (*Adrianichthys*, *Xenopoecilus*)

Family Horaichthyidae (*Horaichthys*)

Superfamily Cyprinodontoides

- Family Cyprinodontidae (*Fundulus*, *Aplocheilus*)
- Family Goodeidae (*Goodea*)
- Family Jenynsiidae (*Jenynsia*)
- Family Anablepidae (*Anableps*)
- Family Poeciliidae (*Poecilia*, *Xiphophorus*)
- Suborder Atherinoidei
  - Superfamily Atherinoidea
    - Family Melanotaeniidae
    - Family Atherinidae (*Atherina*)
    - Family Isonidae, new family (*Iso*)
  - Superfamily Phallostethoidea
    - Family Neostethidae (*Neostethus*)
    - Family Phallostethidae (*Phallostethus*)

## The family Oryziatidae

Rosen (1964) erected a new monogeneric family and described the following diagnoses of the family Oryziatidae. Type genus: *Oryzias* Jordan and Snyder, 1906. Diagnoses: The Oryziatidae differ from their closest relatives, the adrianichthyids, in lacking the tremendously enlarged jaws and ethmoideum, in having paired supraoccipital processes (rather than a single median process), and in having the inferior pharyngeal bone distinctly separated (rather than united), and from all cyprinodontoids as follows: autopalatine usually capped by sesamoid bone; pterygoquadrate cartilage forming dorsal process; lower end of premaxilla not hooked or trapezoidal, situated below maxilla rather than between maxilla and dentary bone; first pleural rib on third vertebra; supracleithrum wanting; pelvic bones with upright lateral spurs and not joined midventrally; hypochordal musculature present on caudal fin.

Composition: Rosen listed following seven species of a single genus, *Oryzias*: *O. latipes* (Temminck and Schlegel), *O. melastigma* (McClelland), *O. celebensis* (Weber), *O. timorensis* (Weber and de Beaufort), *O. javanicus* (Bleeker), *O. curvinotus* (Nichols and Pope), and *O. minutillus* Smith. To these, *O. luzonensis* (Herre and Ablan) may be added. Besides these, Turner (1965) mentioned *O. matenensis* (Aurich), and *O. marmoratus* (Aurich) from the Celebes.

Probably not all these nominal species are valid, since some nominal species are different only in the anal fin-ray frequency.

## The genus *Oryzias*

The following is the diagnoses of the genus *Oryzias* described by Jordan and Snyder (1906), basing on *O. latipes* which has previously been known as *Aplocheilus latipes*.

Body elliptical in form, compressed, covered with large scales; mouth small, with two rows of small, simple, pointed teeth; *no teeth on vomer*\*1; gill-opening not restricted above; intestinal canal short, about as large as body; peritoneum black. Dorsal fin short, inserted above middle of anal; anal very long seventeen to twenty rays; caudal fin truncate. *Sexes similar*\*2 *except color*; anal fin not modified in the male. \*1 Kulkarni(1948) first showed that *os vomer* is absent in *Oryzias melastigma*. \*2 Sexual

dimorphism is prominent. See Chap. 8.

## The species *Oryzias latipes*

The following description by Oshima (1919) based on a specimen of *Oryzias latipes* collected from Shori, Formosa is cited here as the diagnoses of the species since it is very precise and correct excepting two words starred and daggered.

Head 4 in length (body length divided by head length is 4); depth 4.5; depth of caudal peduncle 9.5; eye 2.5 in head (head length divided by eye diameter is 2.5); interorbital space 2; snout 4; D.6; A.18; P.9; V.5; thirty one scales in a lateral series; five branchiostegals.

Posterior half of the body compressed, becoming broader anteriorly, highest in front of the anal; head flattened; interorbital space broad; snout shorter than the diameter of eye, broadly rounded anteriorly; mouth anterior, transverse; lower jaw slightly projecting, each jaw with two rows of minute pointed teeth, those on the posterior row smaller; vomer\*1 smooth; thirteen short, pointed gill-rakers on the first arch; eyes very large, anterior and superior.

Dorsal fin short, on the posterior half of body, its origin above the posterior two thirds of anal, its height equal to the distance between tip of snout and posterior margin of orbit; pectoral inserted on the median line of body; the ventral small, reaching vent; base of the anal very long, its posterior end opposite to that of the dorsal, anterior ray longest; tip of the caudal fin *rounded*.\*2

Top and sides of head, throat, and chin naked; body covered with cycloid scales, lateral line absent.

Color in formalin pale gray above, lower parts silvery; a black longitudinal streak from the nape to the origin the dorsal; sides of body with a faint dusky stripe along the middle line, top of head dark; the edges of scales dusky; fin-rays of the ventral and anal dotted with minute black spots; all the fins whitish; peritoneum black. Length of body 28 mm.

Habitat: The present species is very common in rice-fields and pools on the island.

\*1 Vomer is absent in *Oryzias* in reality.

\*2 The caudal fin is almost truncate, strictly, however, it is incipiently lobed.

## Change of nomenclature of the medaka

The medaka was first described as *Poecilia latipes* by Temminck and Schlegel in 1846 (Siebold's Fauna Japonica, Poiss., P.224, Pl.102, Fig. 5). Gŷnther changed it as *Haplochilus latipes*. Jordan and Snyder (1901) described it as *Aplocheilus latipes* but later they separated it from *Aplocheilus* and erected a new genus *Oryzias*. They regarded *Oryzias* as having no teeth on vomer\*1 while *Aplocheilus* possesses teeth on it.

Myers (1931) placed the medaka in the tribe Aplocheilini of the subfamily Fundulinae in the family Cyprinodontidae. He stated that the chief character of fishes of the tribe is the non-protractile premaxillae. The pectoral fin are set high and pseudobranchiae and vomerine teeth are never present. The species range from Japan and Central China south to Celebes and Timor and west to Southern India. A single genus, *Aplocheilus*, of which *Oryzias* is a synonym\*2. Smith, (1945) pointed out that the genus known as *Panchax* is a synonym of *Aplocheilus* McClelland and *Aplocheilus* Weber and de Beaufort is a synonym of *Oryzias* Jordan and Snyder. He described *Aplocheilus panchax* (Hamilton) and *Oryzias minutillus* n. sp. from Thailand.

According to him, the two genera may be distinguished by the following characters:

- a. Upper jaw protractile; mouth moderate size with its corners abruptly bent downward; vomer toothed; pseudobranchiae present; branchial membranes free from each other and from isthmus; pectoral fins with their upper base at or below longitudinal axis of body . . . . . *Aploc*
- b. Upper jaw not protractile; mouth small with its corners obtusely bent downward; vomer toothless; no pseudobranchiae; branchial membranes united across isthmus; pectoral fins with upper base well above longitudinal axis of body . . . . . *Oryzia*

The correct scientific name of the medaka is *Oryzias latipes* (Temminck and Schlegel).

From Jordan and Snyder (1906) onwards, all taxonomists stated that *Oryzias* has toothless vomer while *Aplocheilus* has toothed vomer. Kulkarni (1948) has made a precise osteological study of Indian killifishes and found that vomer is absent in both *Oryzias melastigma* and *Horaichthys setnai* while *Aplocheilus lineata* possesses toothed vomer.

\*1 Vomer is absent in *Oryzias* in reality.

\*2 Now, the two genera belong to the different super-families.

## Geographical distribution of species belonging to the Genus *Oryzias*

All the species of the genus *Oryzias* are distributed in India, South Asia, the Indo-Australian archipelago and the Far East. Their habitats are widely ranged from tropical, subtropical, and temperate regions as shown in Figure 2-4 and in the following lines.

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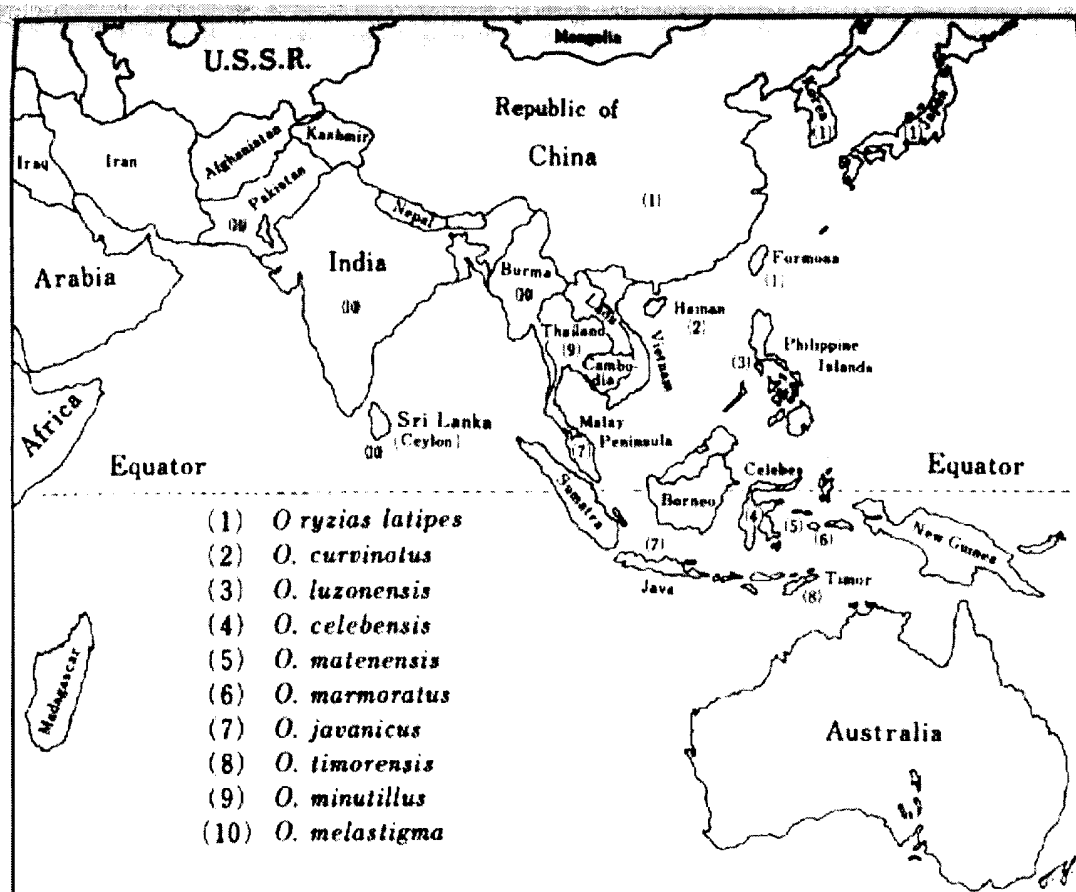


Fig. 2-4. A

zoogeographical map showing distribution of species of the Genus *Oryzias*. Original.

- ( 1 ) *O. latipes* (Temminck and Shlegel): Japan, Korea, Formosa, and China
- ( 2 ) *O. curvinotus* (Nicols and Pope): The island of Hainan.
- ( 3 ) *O. luzonensis* (Herre and Ablan): Luzon in the Philippines.
- ( 4 ) *O. celebensis* (Weber): The Celebes.
- ( 5 ) *O. matenensis* (Aurich): The Celebes.
- ( 6 ) *O. marmoratus* (Aurich): The Celebes.
- ( 7 ) *O. javanicus* (Bleeker): The Indo-Malaysian archipelago and Malaya.
- ( 8 ) *O. timorensis* (Weber and de Beaufort): The island of Timor.
- ( 9 ) *O. minutillus* Smith: Thailand.
- (10) *O. melastigma* (McClelland): India, Western Pakistan, and Sri Lanka (Ceylon).

In the main, all the *Oryzias* species are fresh-water fishes. *O. latipes* and *O. melastigma* inhabit both fresh and brackish water. *O. latipes* is so tolerate slinity that it thrives in tide pools in Korea and Kyushu in Japan.

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